

Coremodified Smaragdyrins; First examples of stable meso substituted Expanded Corrole

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Supporting Information :

1. The ^1H NMR spectra for **5a**, **5b**, **5c**, **5d**, **5e**, **5f**, **6a**, **6b** and **6c**. (16 pages)
2. 2D COSY Spectra for **5a** and **6a** and 2D TCOSY spectra for **6a**. (3 pages)
3. Representative mass spectra for **5a**, **5b**, **5d**, **5e**, **6a**, **6b** and **6c**. (7 pages)
4. The electronic (a) and fluorescence (b) spectra of **5a** (solid line) and **6a** (dashed line in CH_2Cl_2 . Excitation wavelengths are 443nm (**5a**) and 411nm (**6a**). (1 page)
5. Table containing the selected physical data for the new compounds (2 pages)
6. General synthetic procedure (1 page)
7. Table containing the yields of various compounds (1 page)
8. The ORTEP plots showing the crystal structure of **6a** (top: plane view, bottom: side view - phenyl rings are omitted for clarity). (1 page)
9. The ORTEP plot showing two independent molecules of **5a** trapping three solvent methanol molecules with weak interactions in the unit cell. (1 page)
10. The ORTEP plots of Rh(I) Carbonyl complex of **5a**. (top: plane view, bottom: side view) (1 page)
11. Tables of crystal data, structure solution and refinement, atomic coordinates, bond lengths and angles and anisotropic thermal parameters for compound **5a**. (15 pages)
12. Tables of crystal data, structure solution and refinement, atomic coordinates, bond lengths and angles and anisotropic thermal parameters for compounds **6a**. (9 pages)

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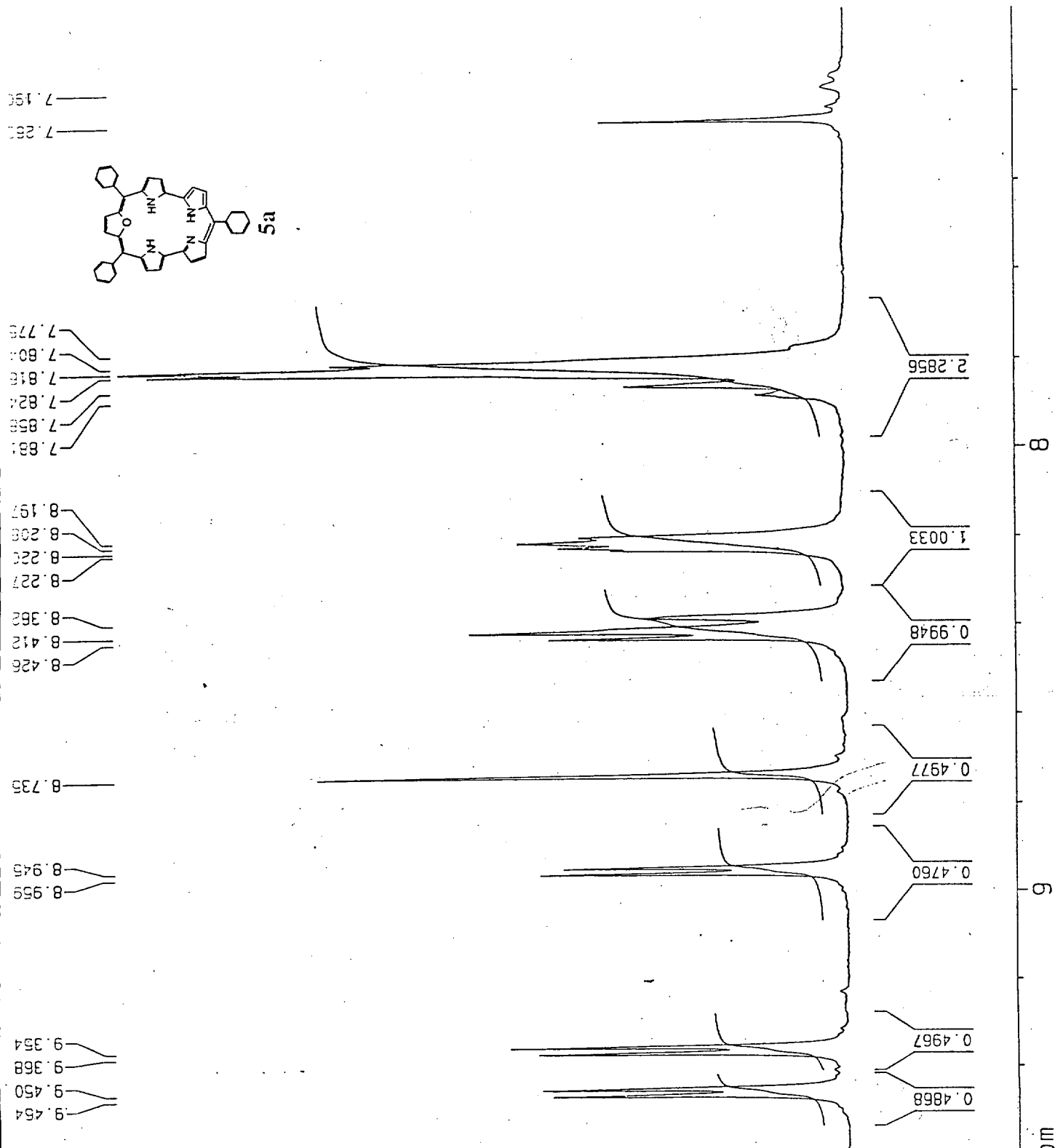
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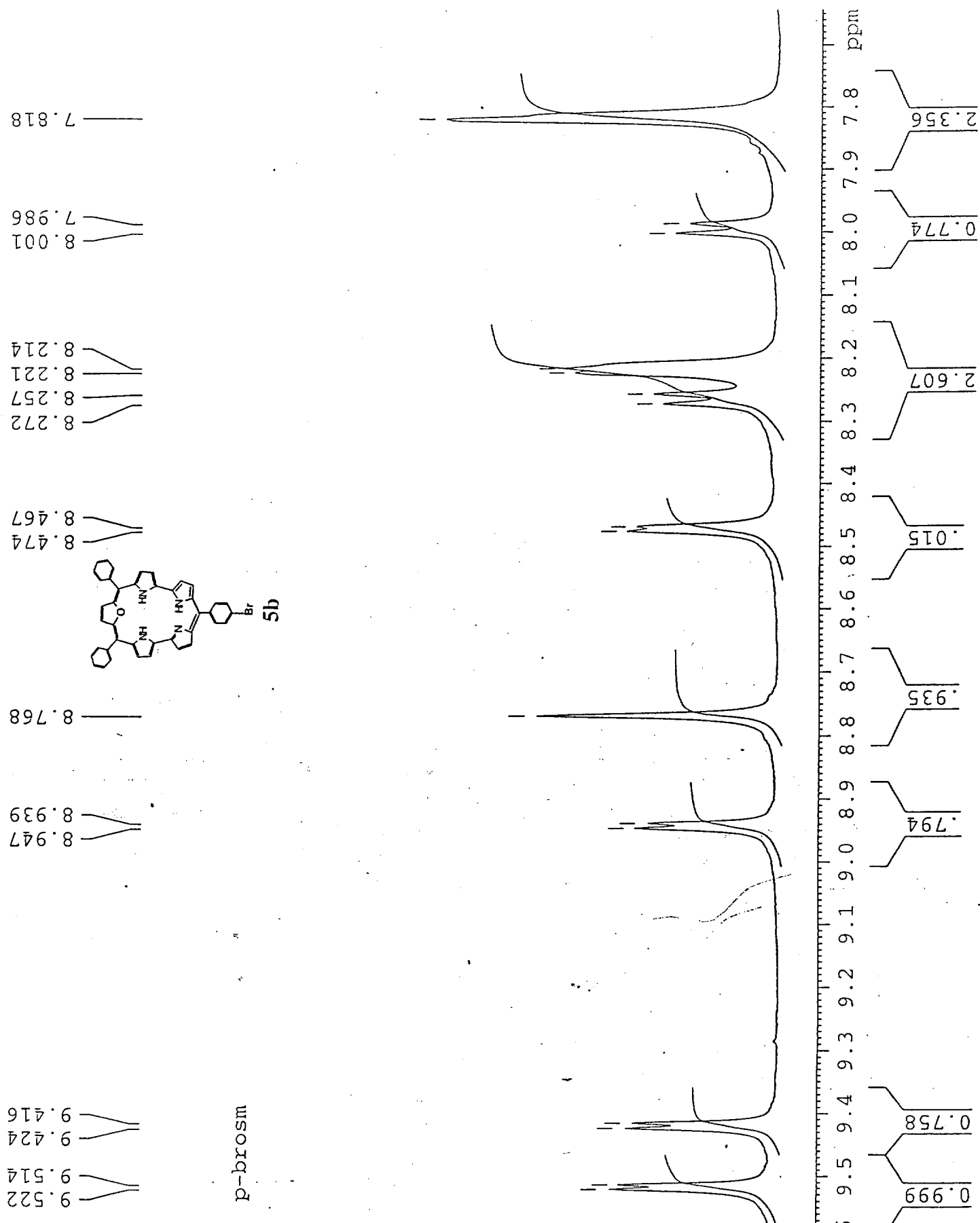
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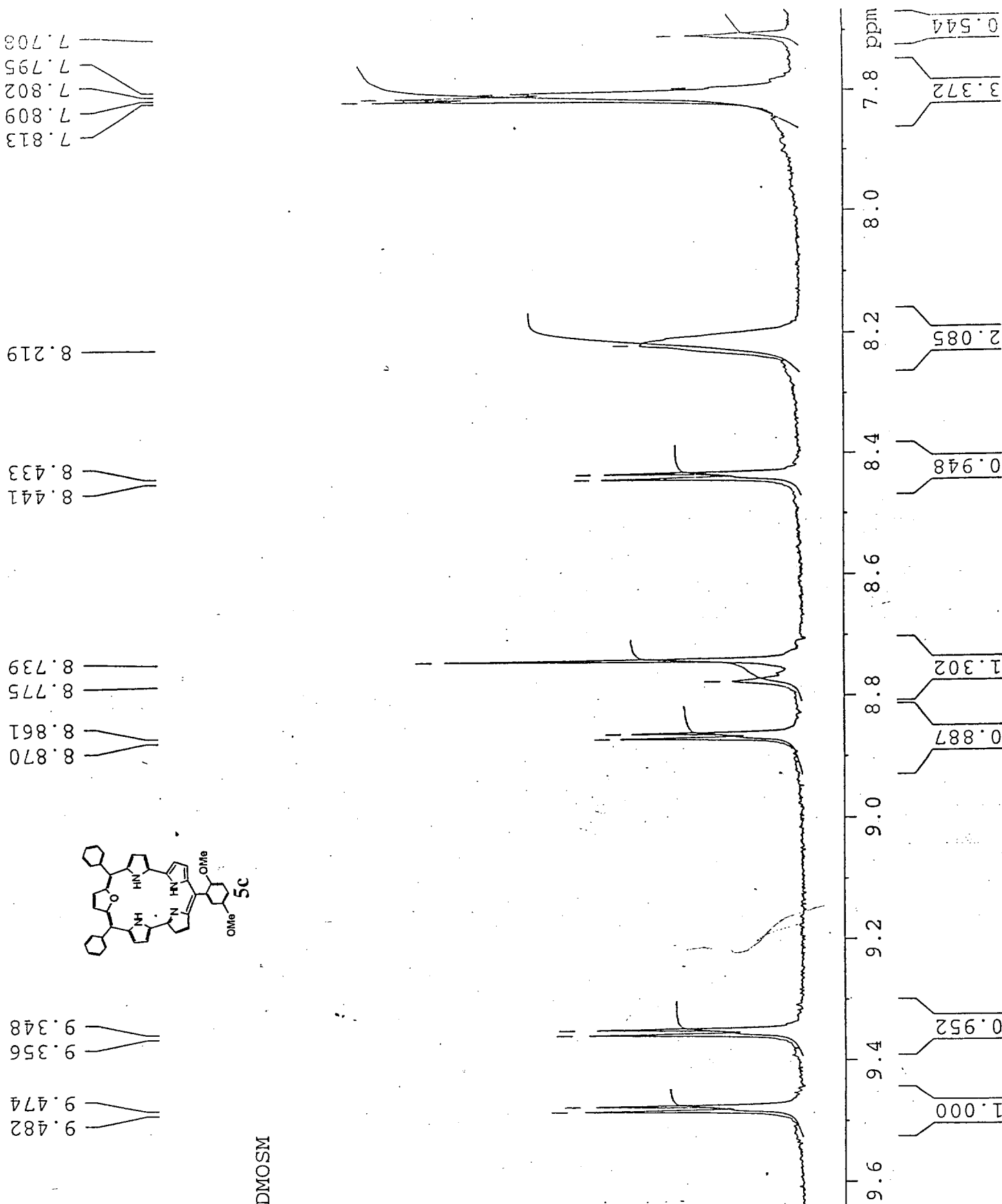
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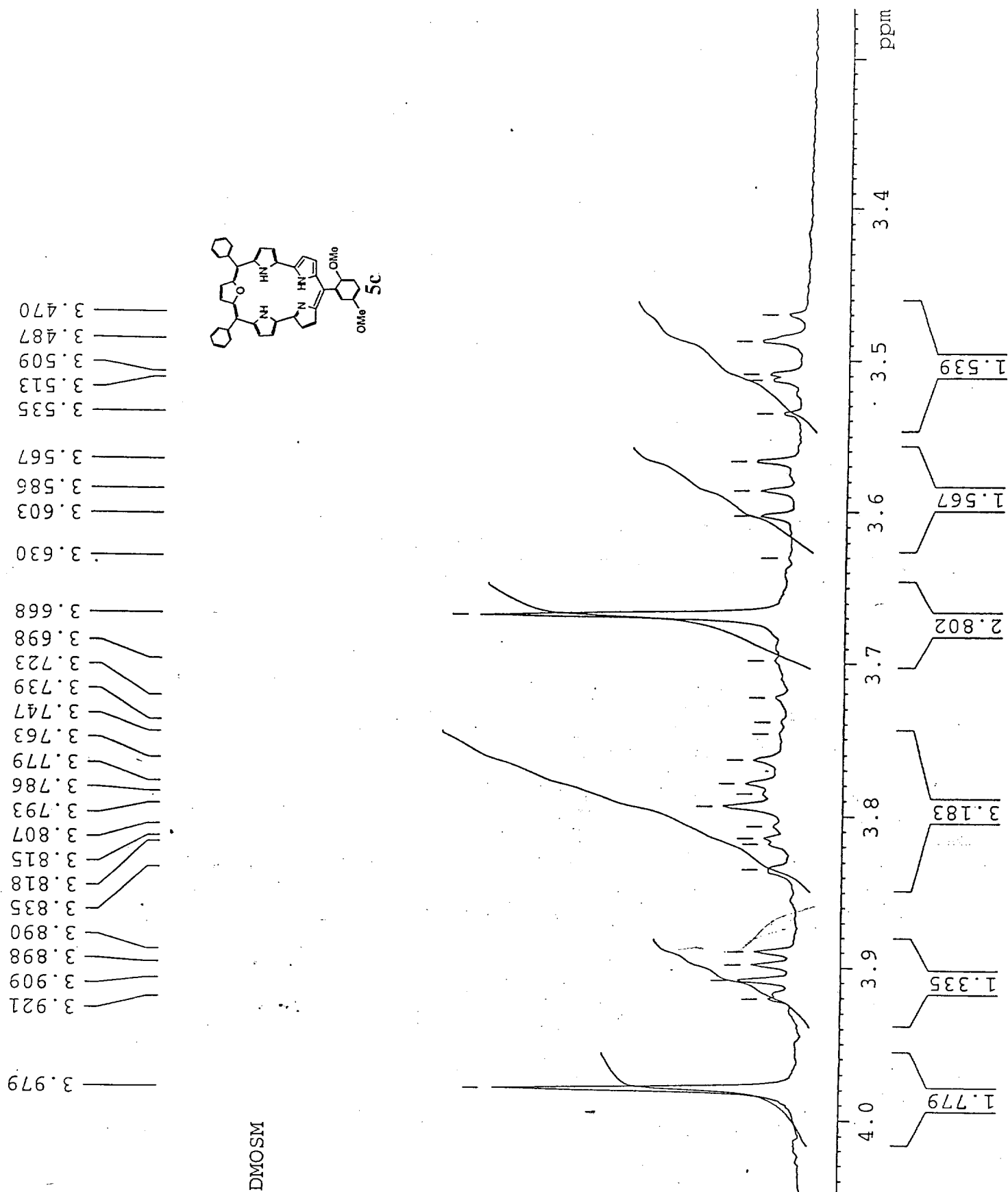


¹H NMR (300 MHz) spectrum of 5a in CDCl₃.

ppm







¹H NMR (500 MHz) spectrum of 5c in CDCl₃.

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7.793

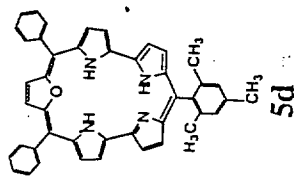
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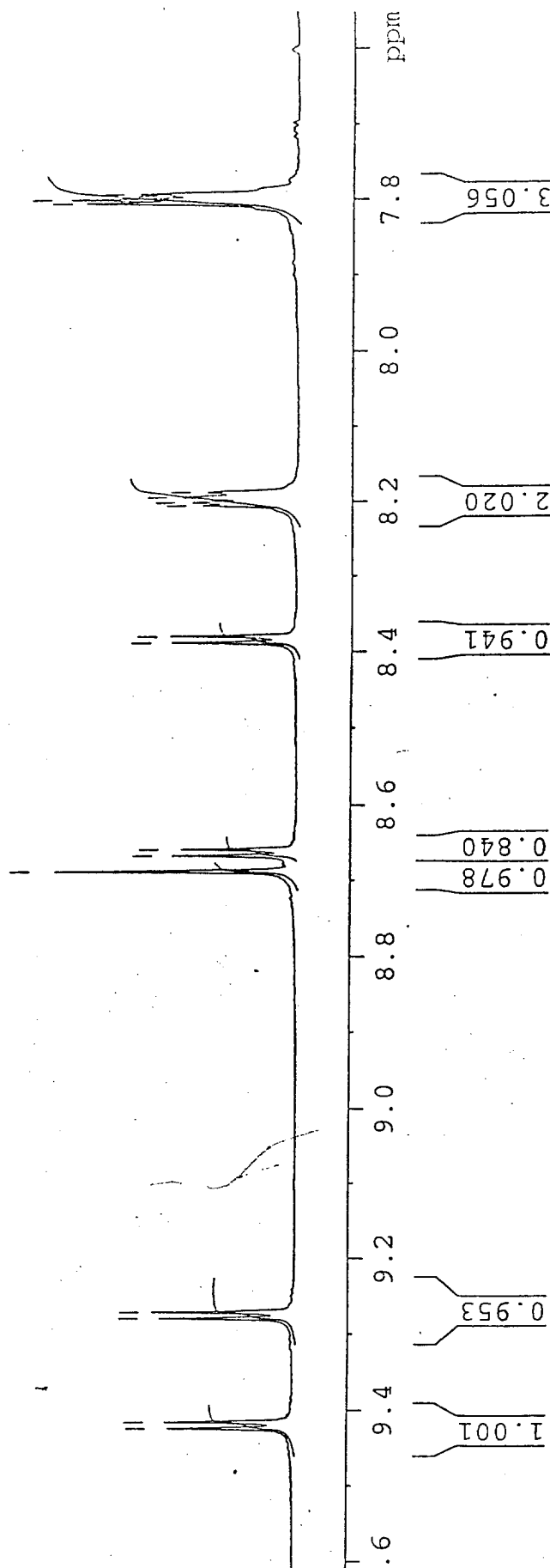
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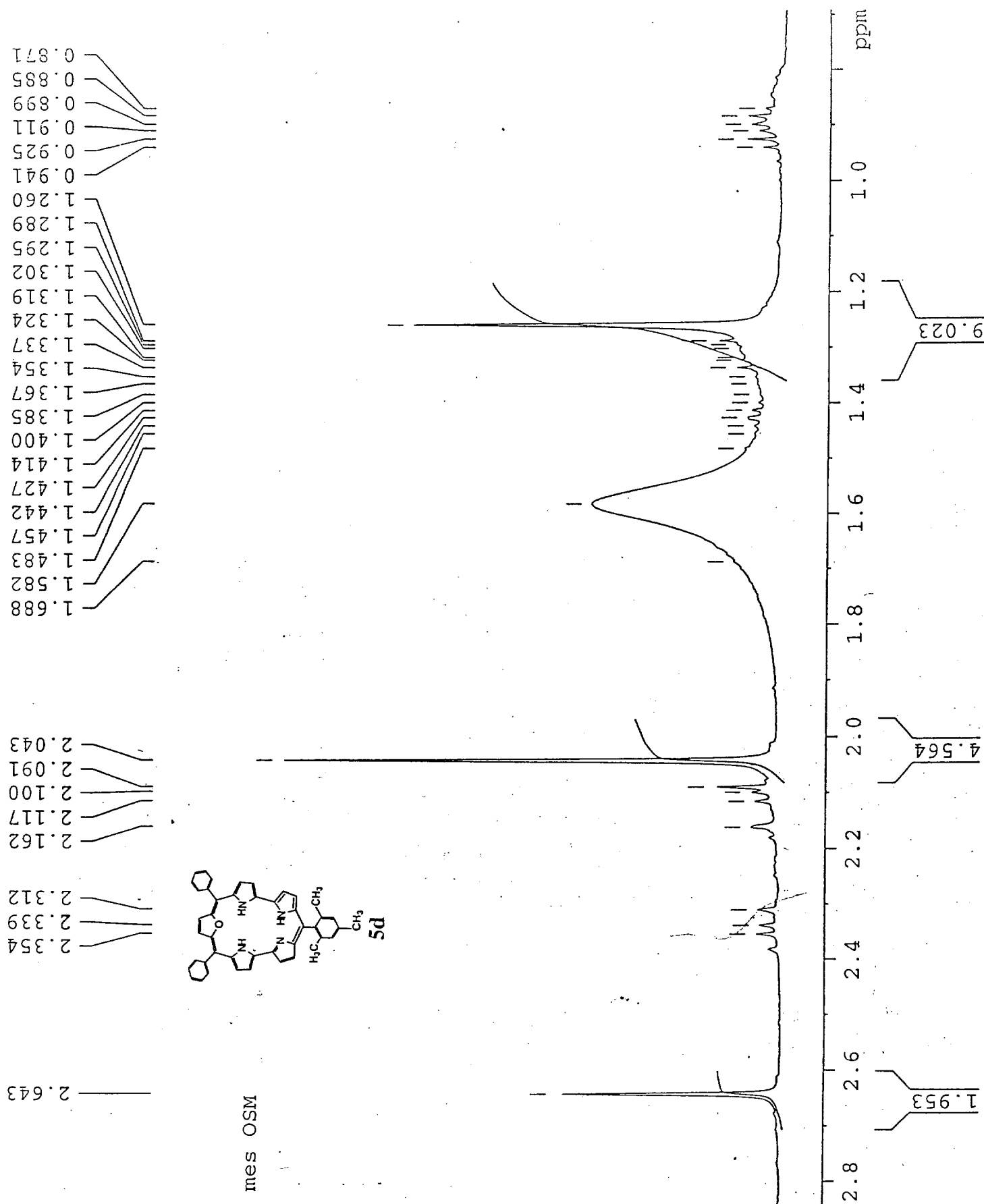
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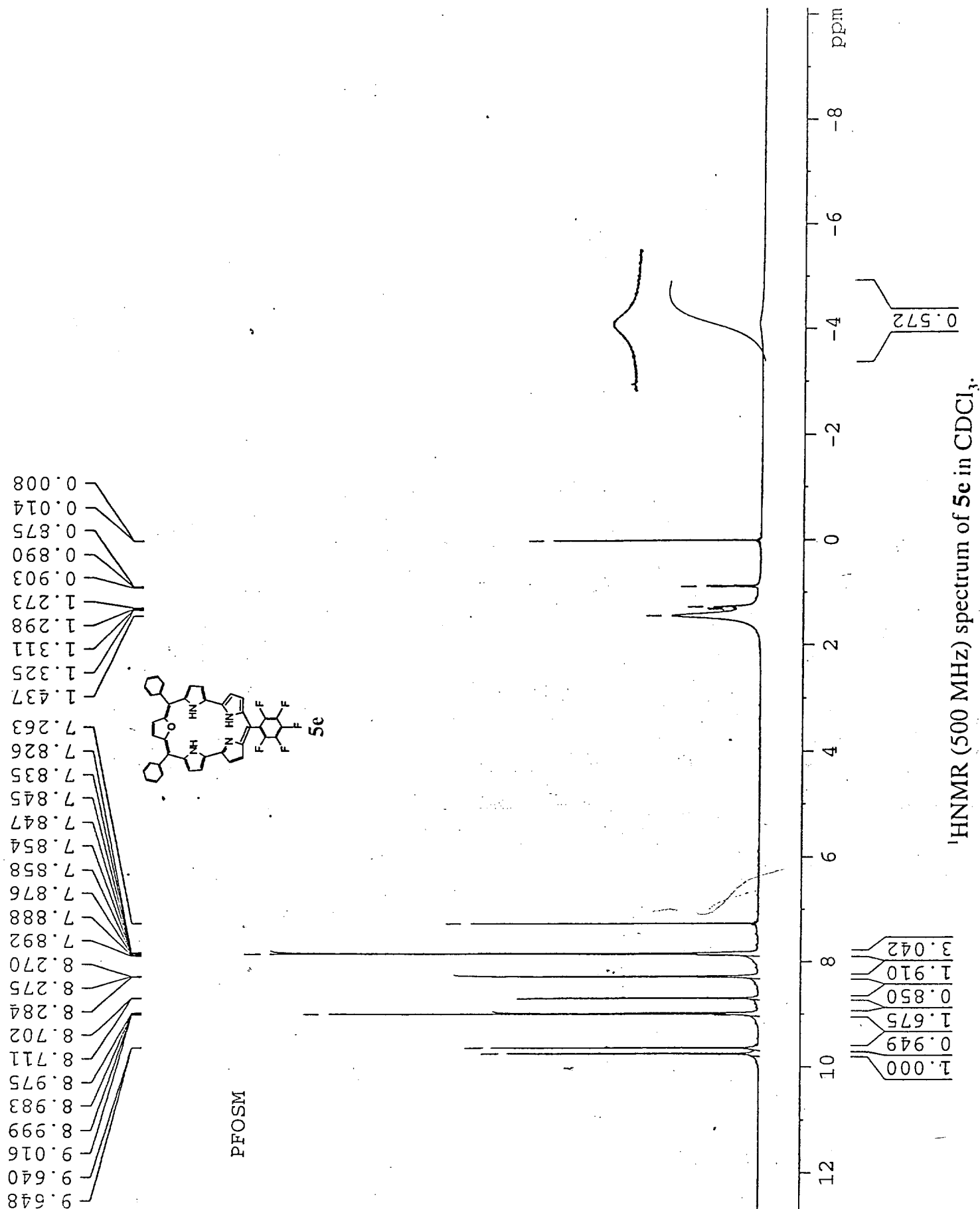
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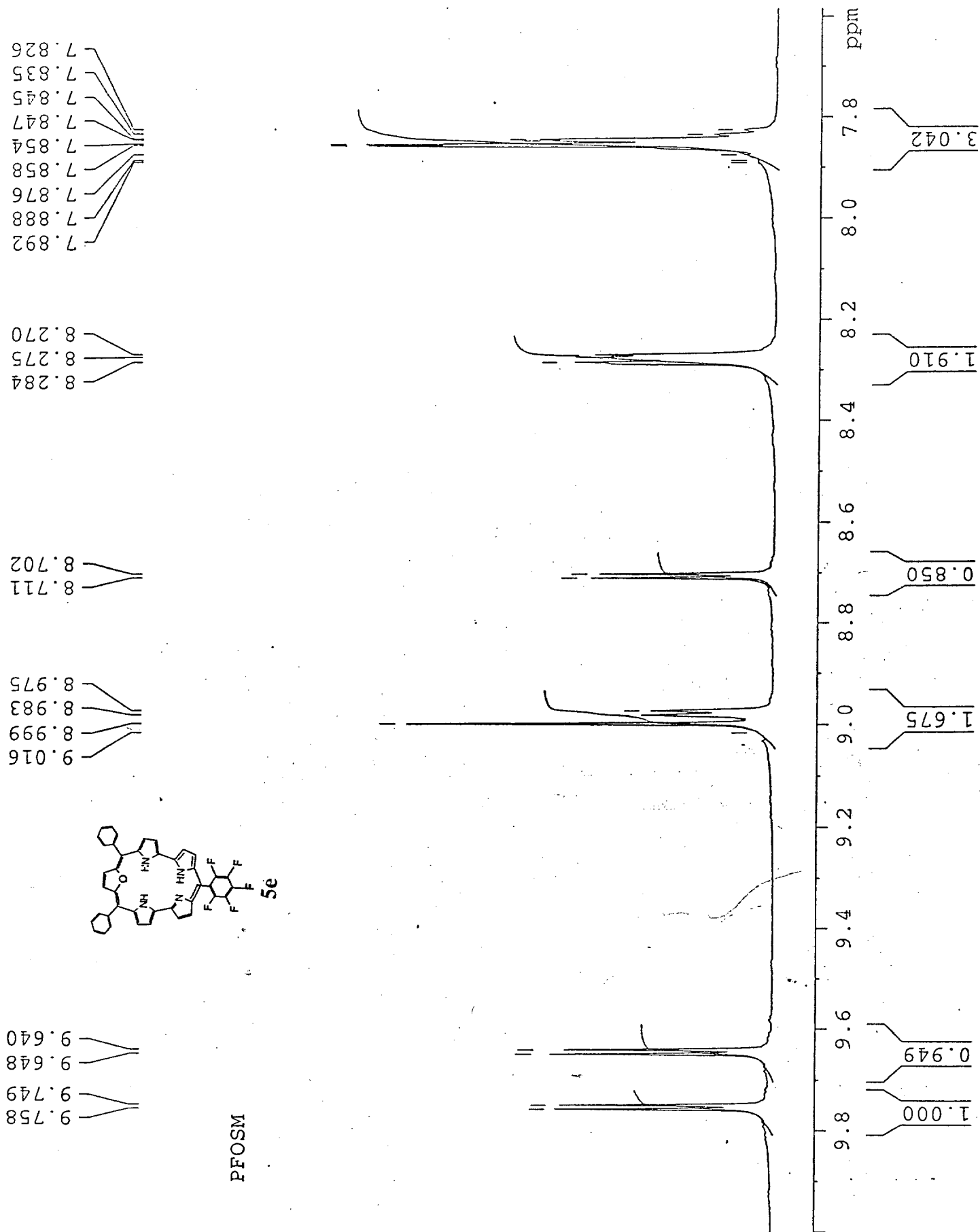


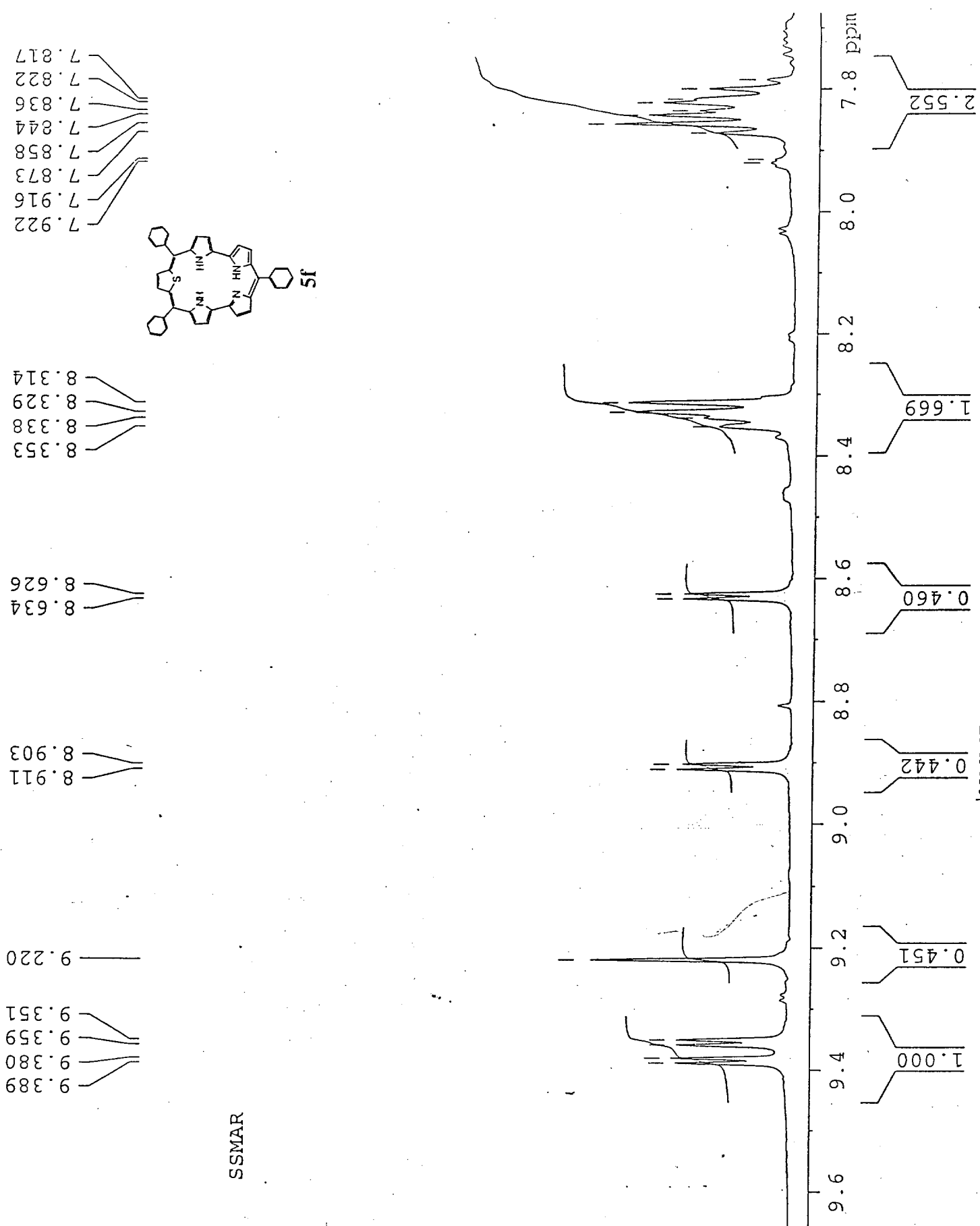
mes OSM











SSMAR

510

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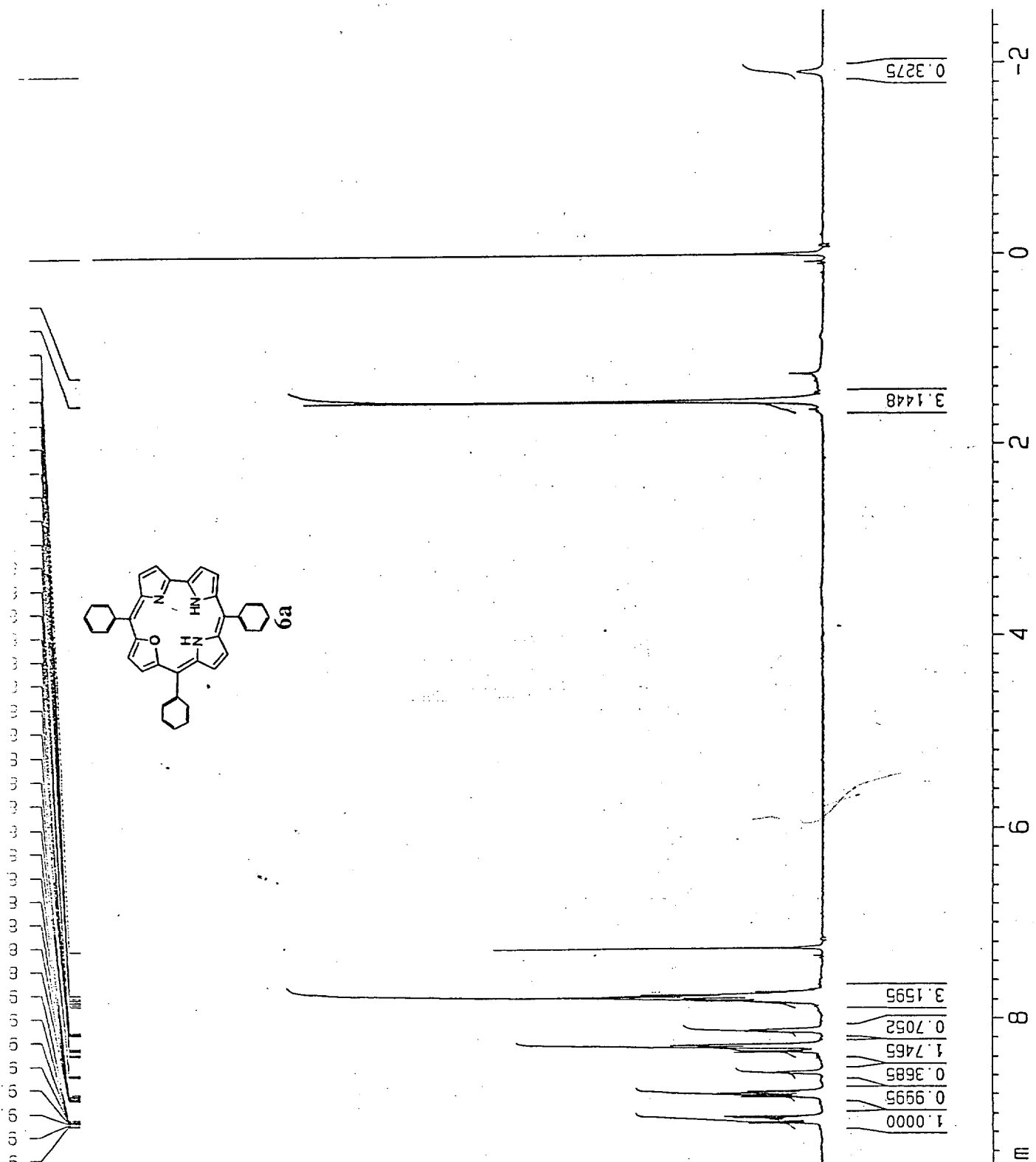
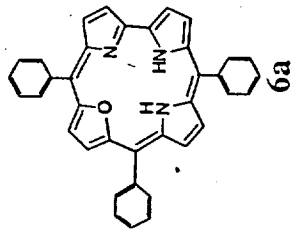
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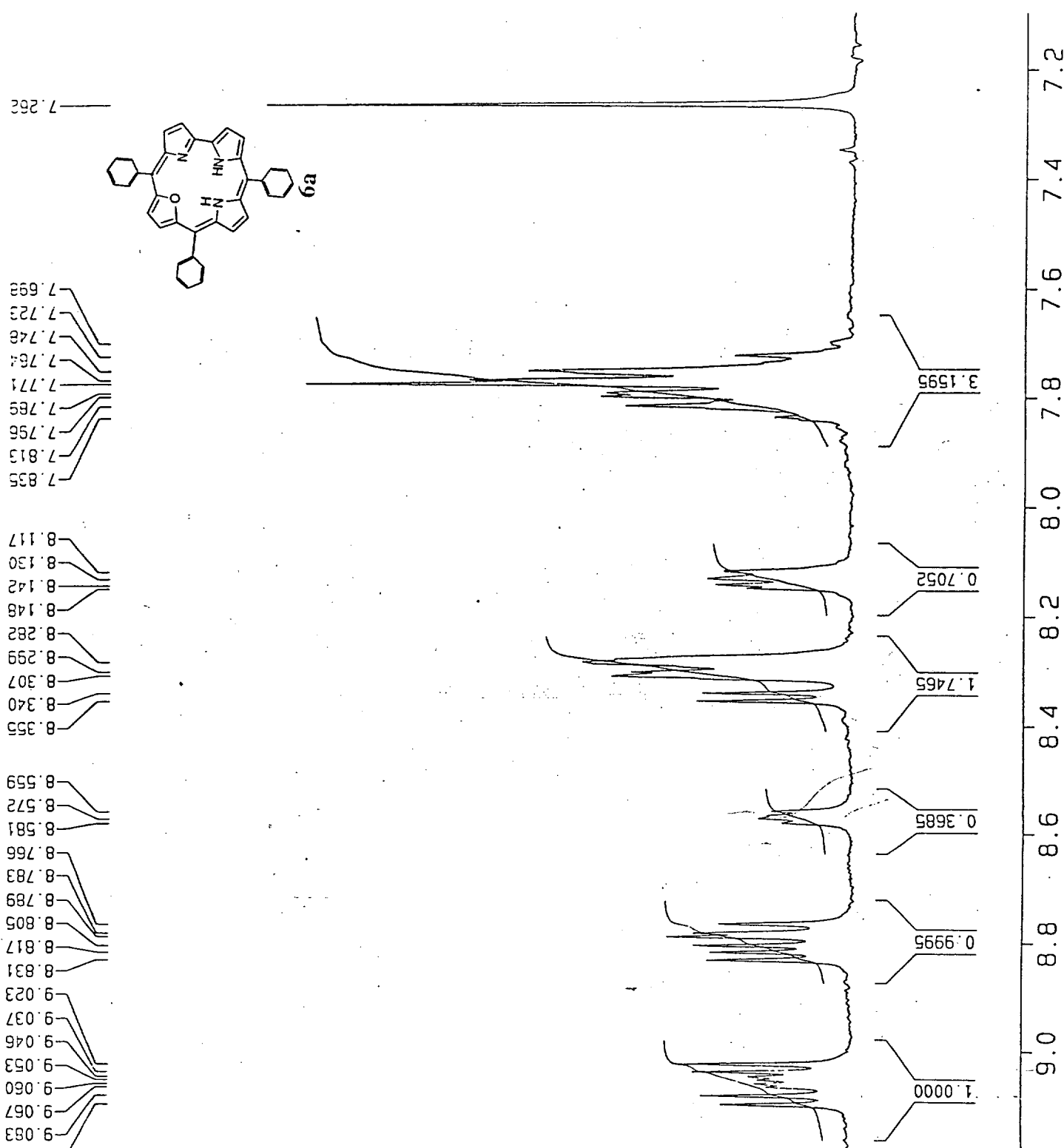
¹H NMR (300 MHz) spectrum of 6a in CDCl₃.

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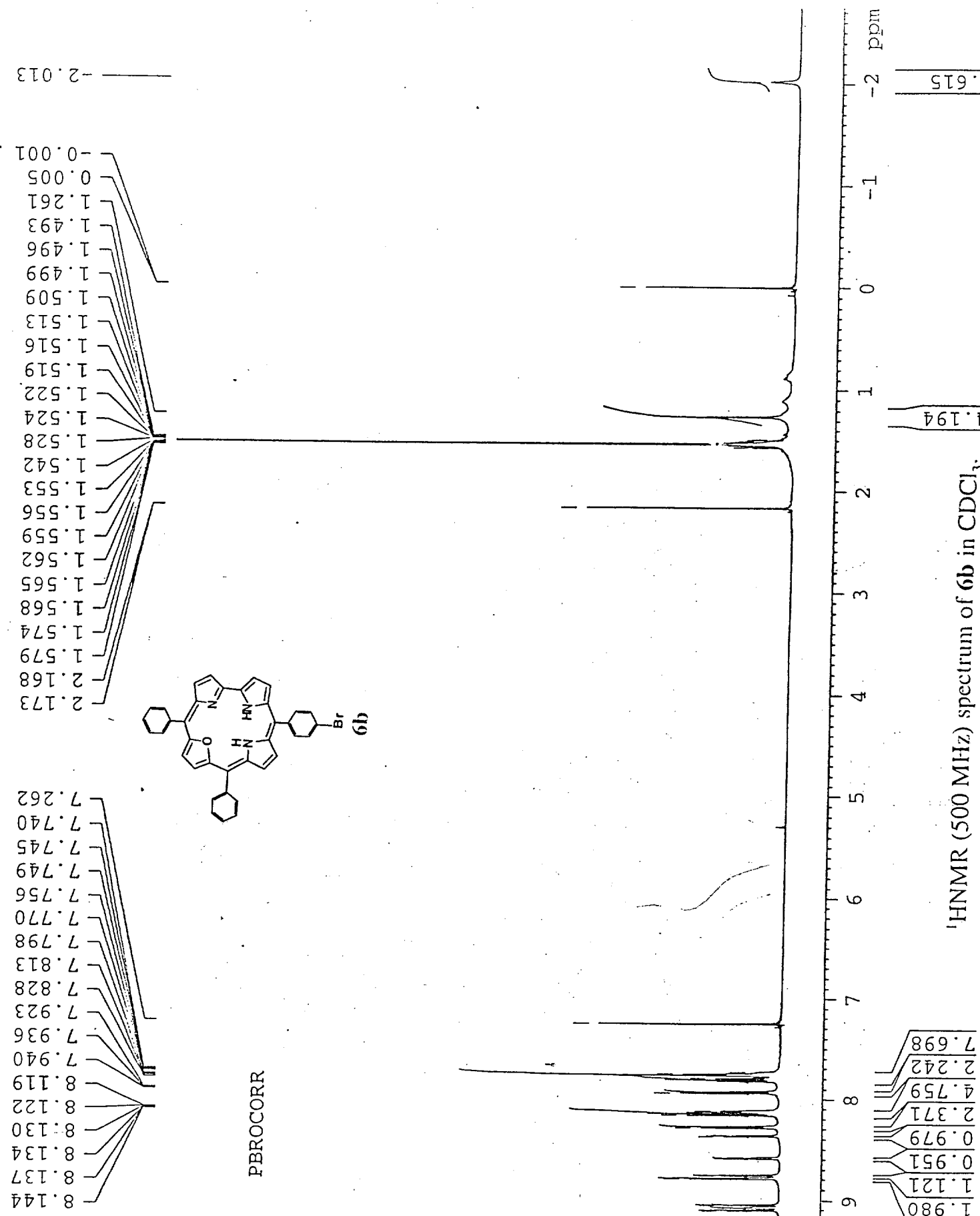
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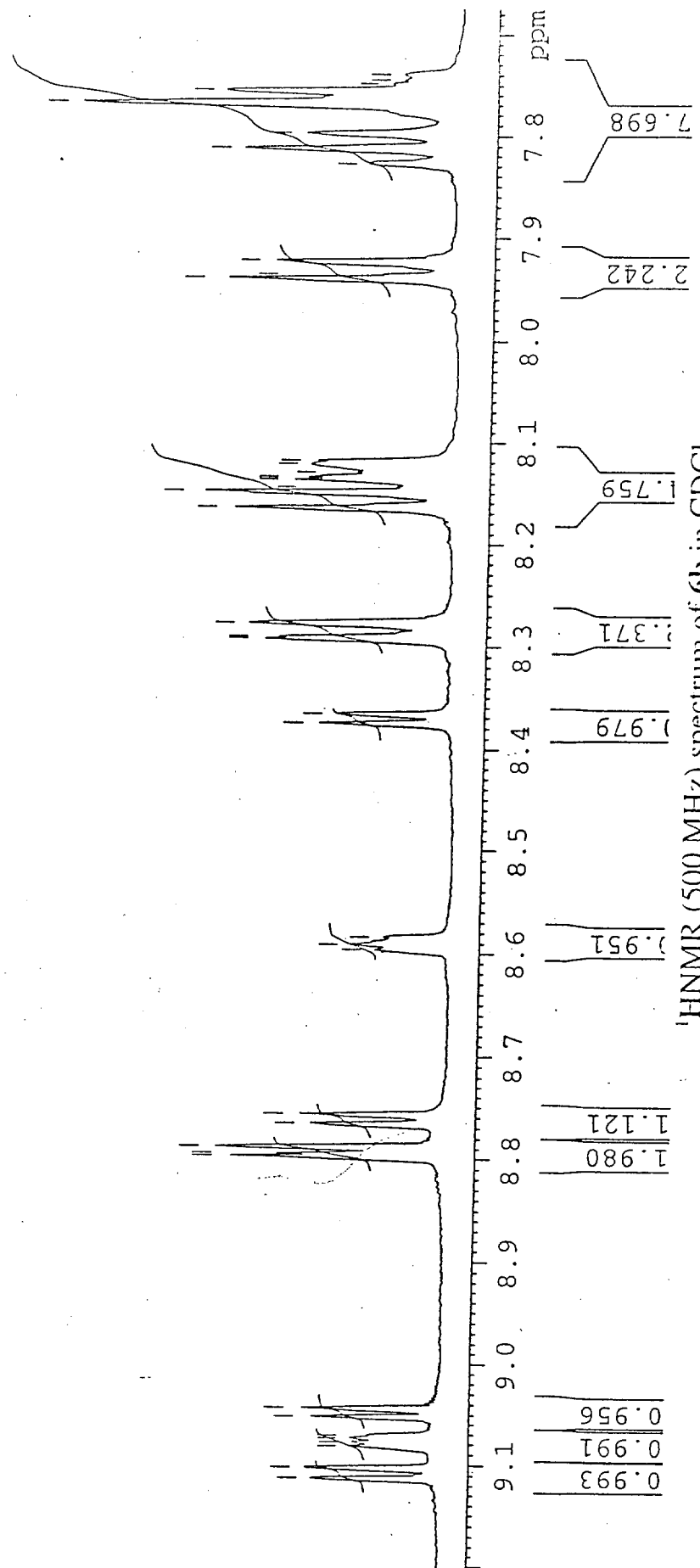
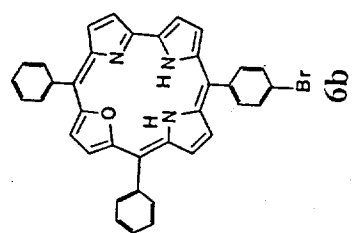
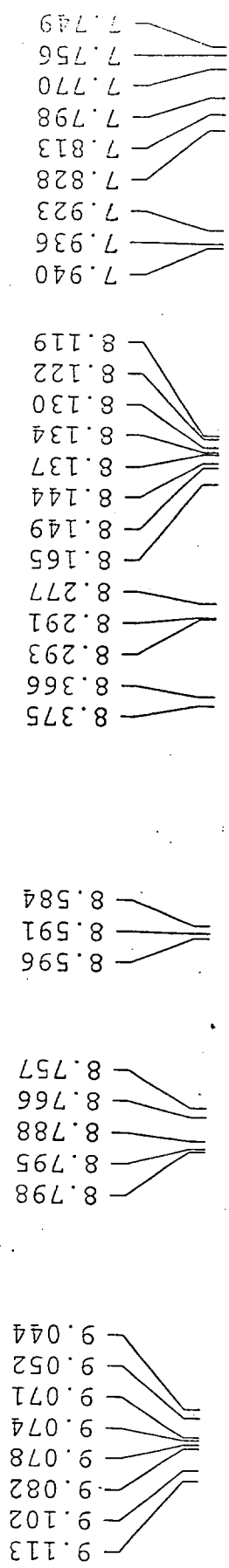
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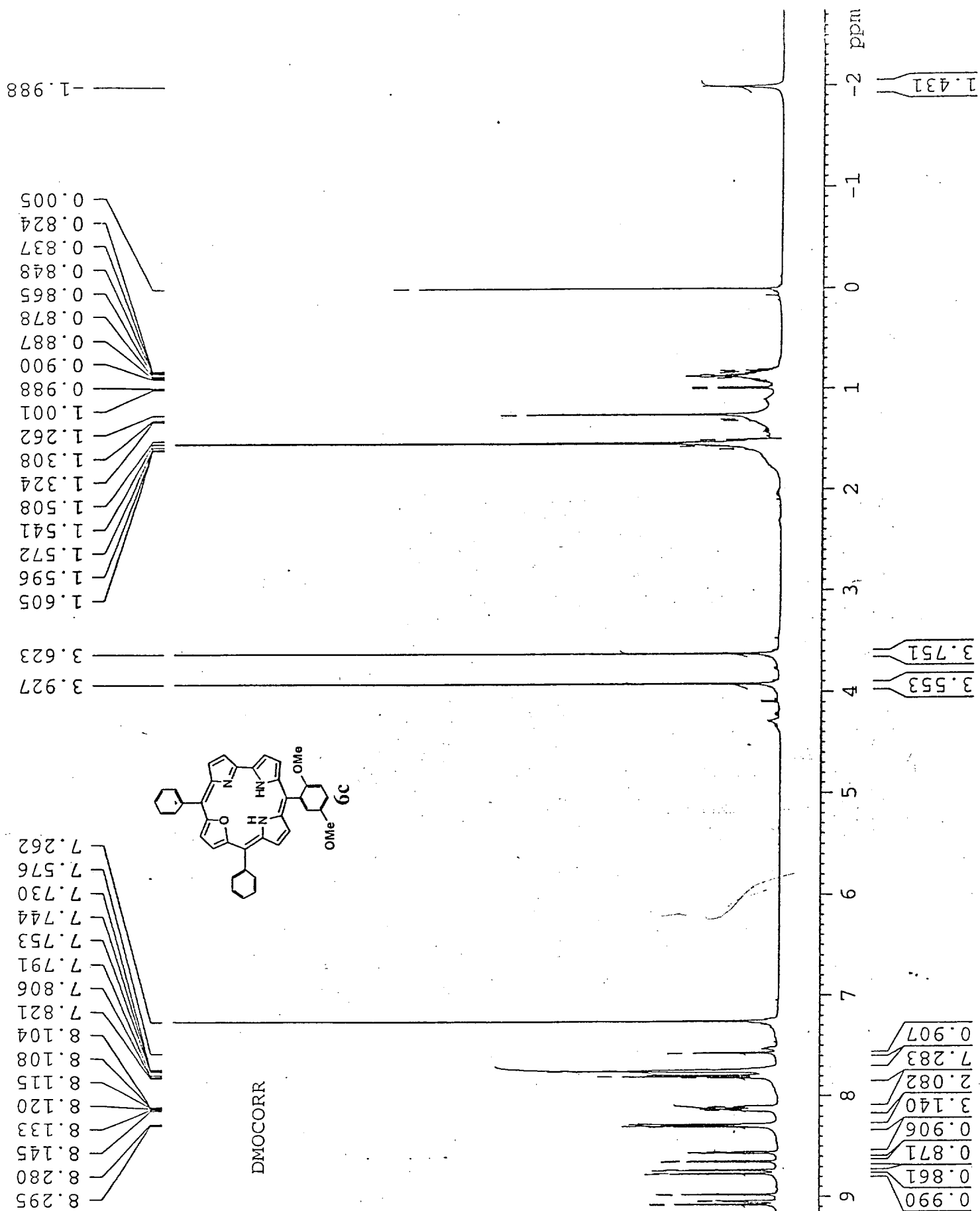


¹H NMR (300 MHz) spectrum of 6a in CDCl₃.

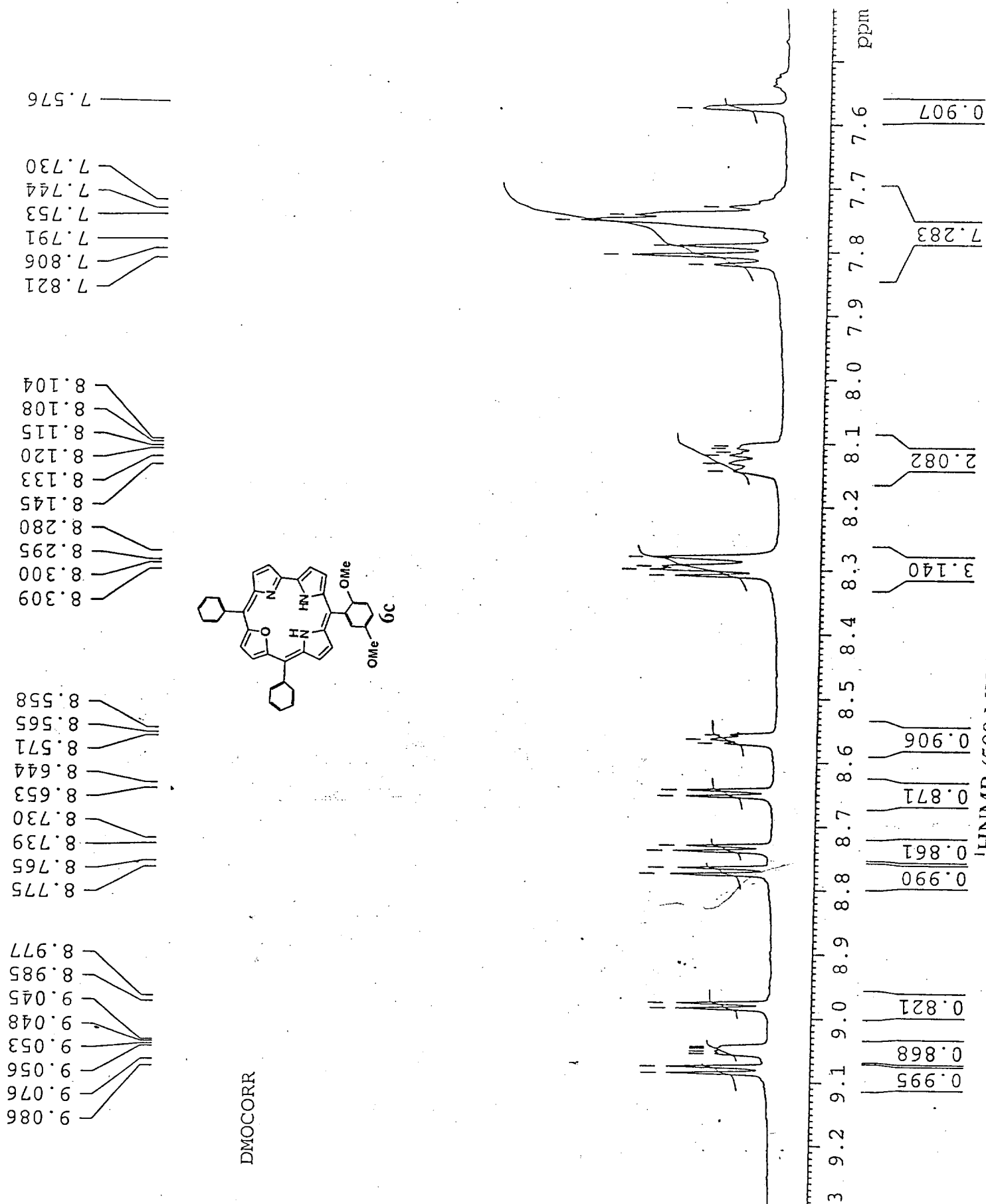


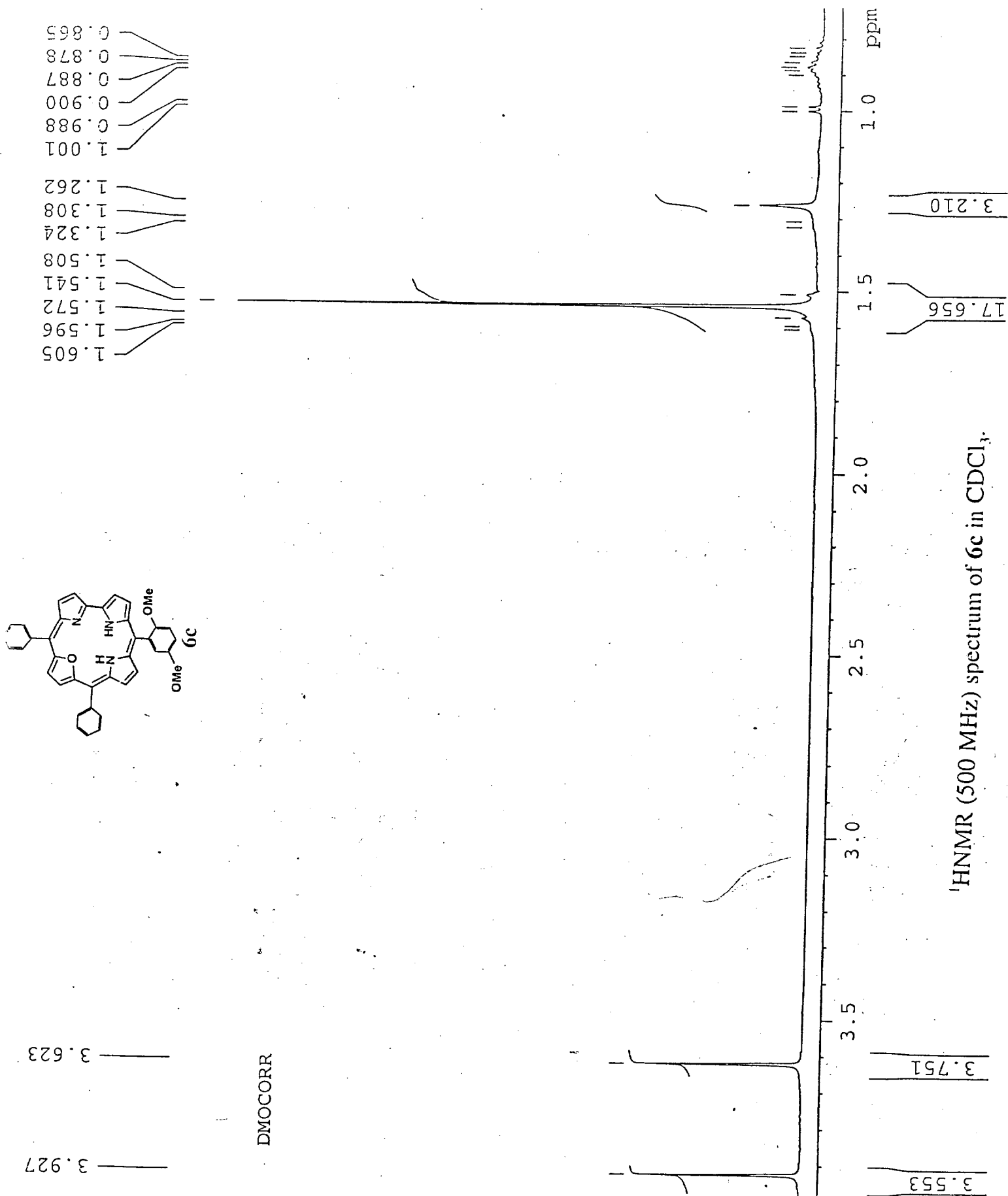
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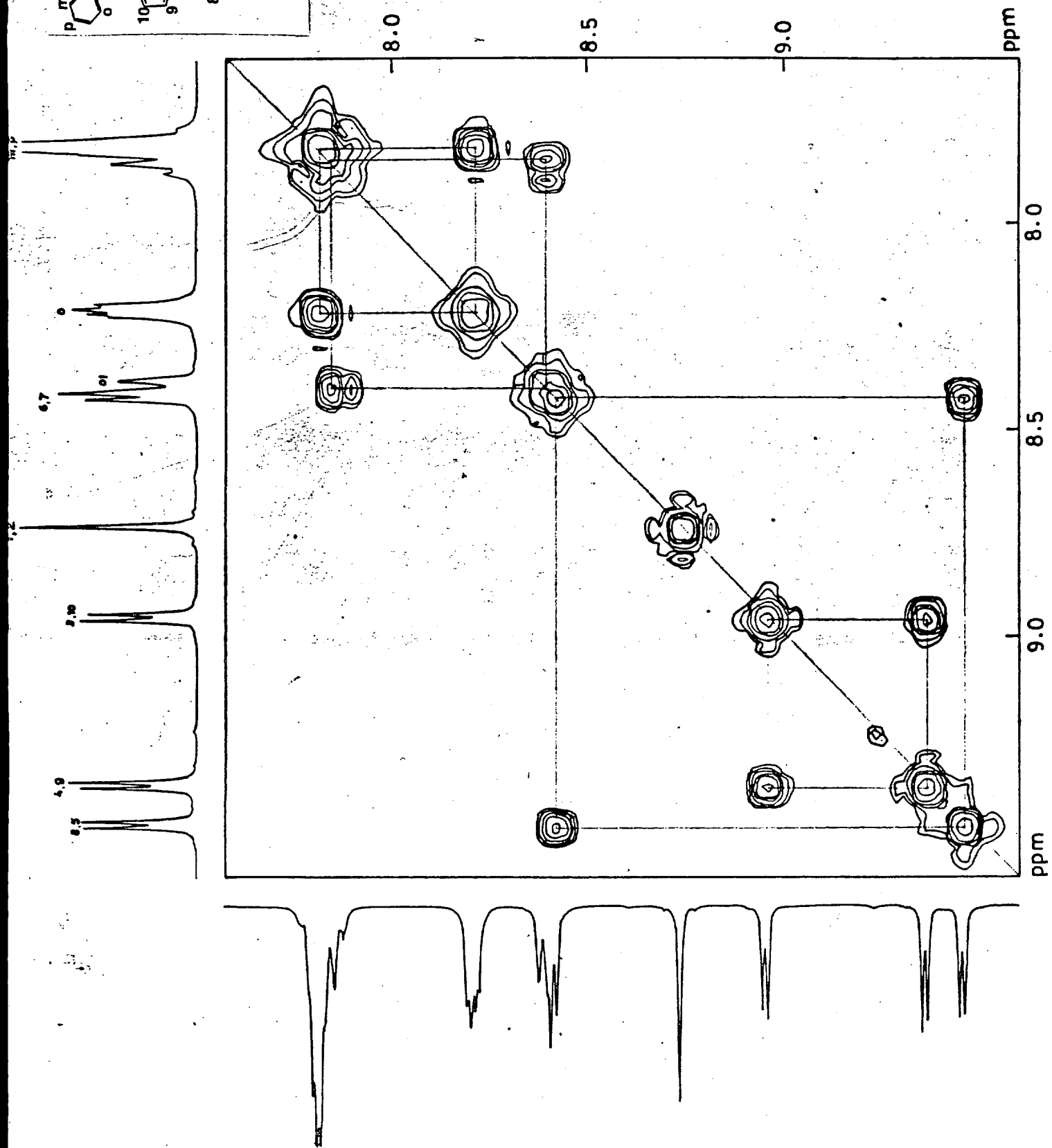
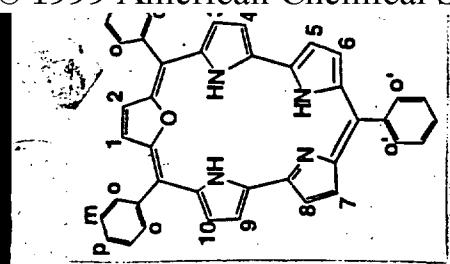




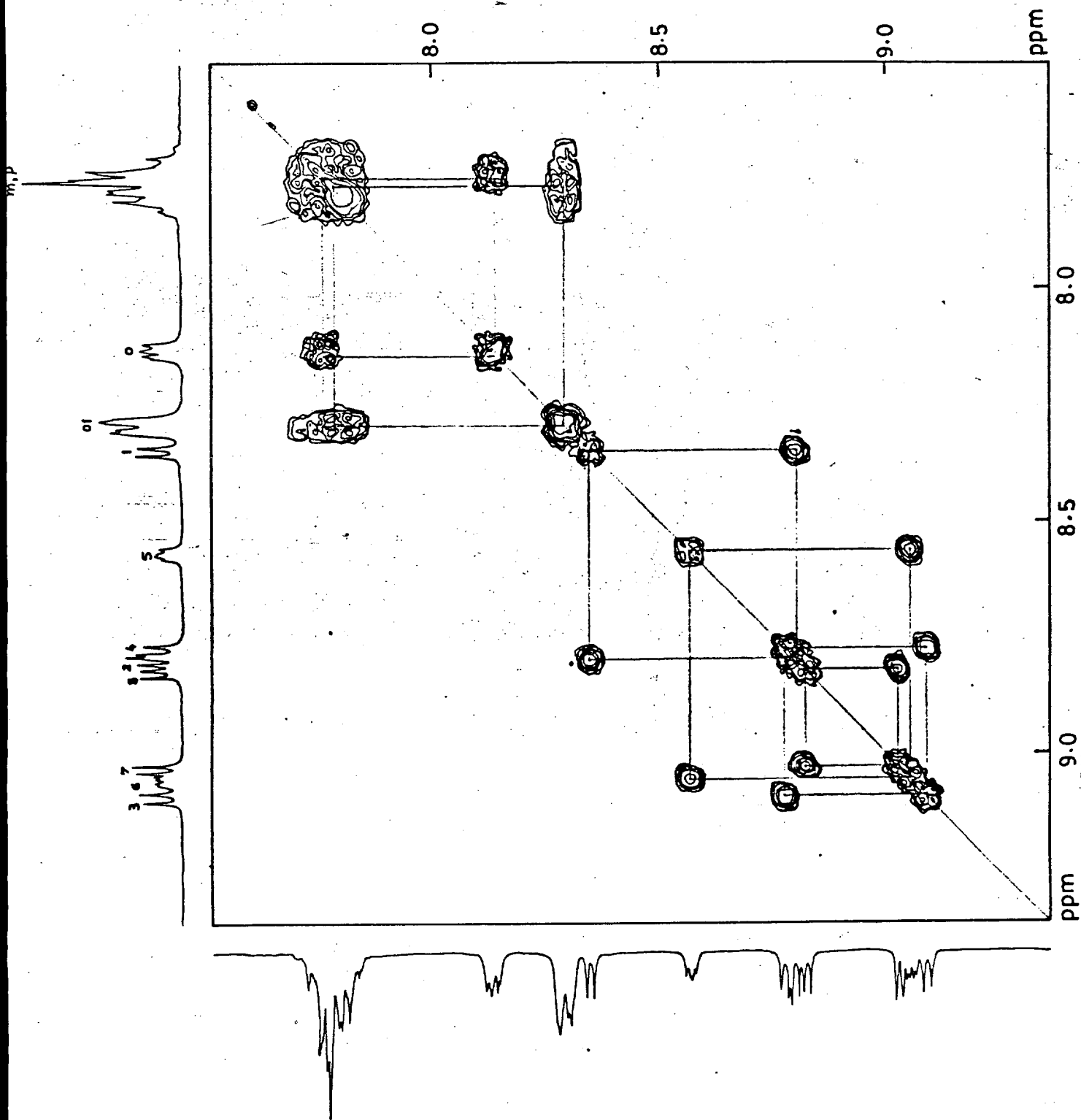
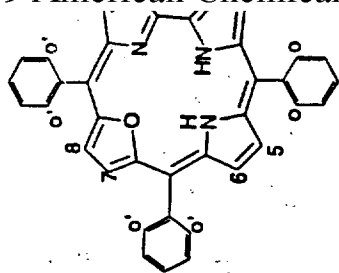
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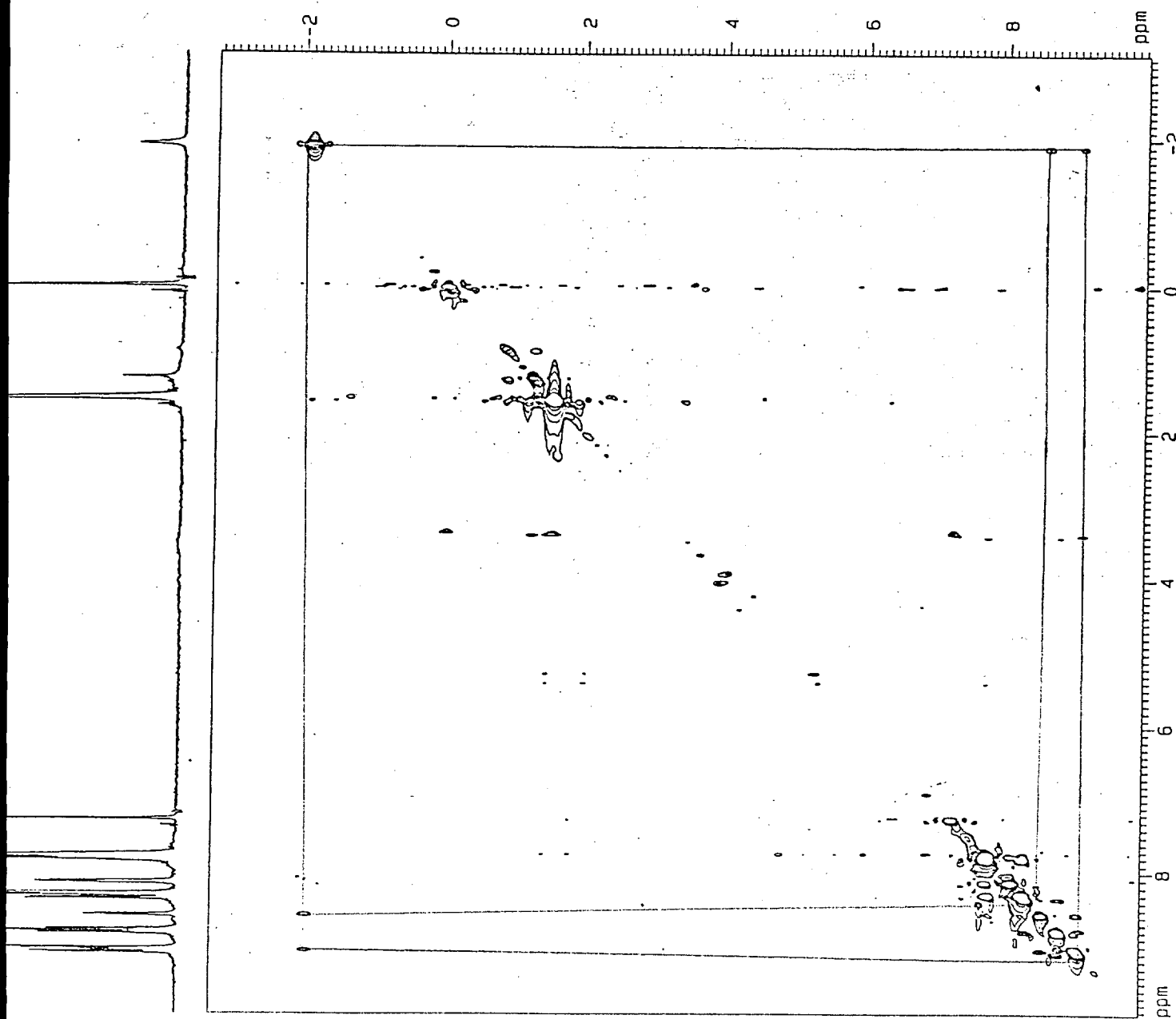
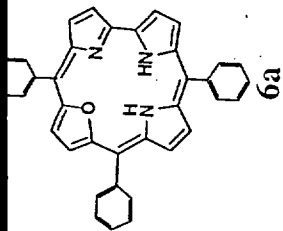




2D-COSY (300 MHz) spectrum of 5a in CDCl₃



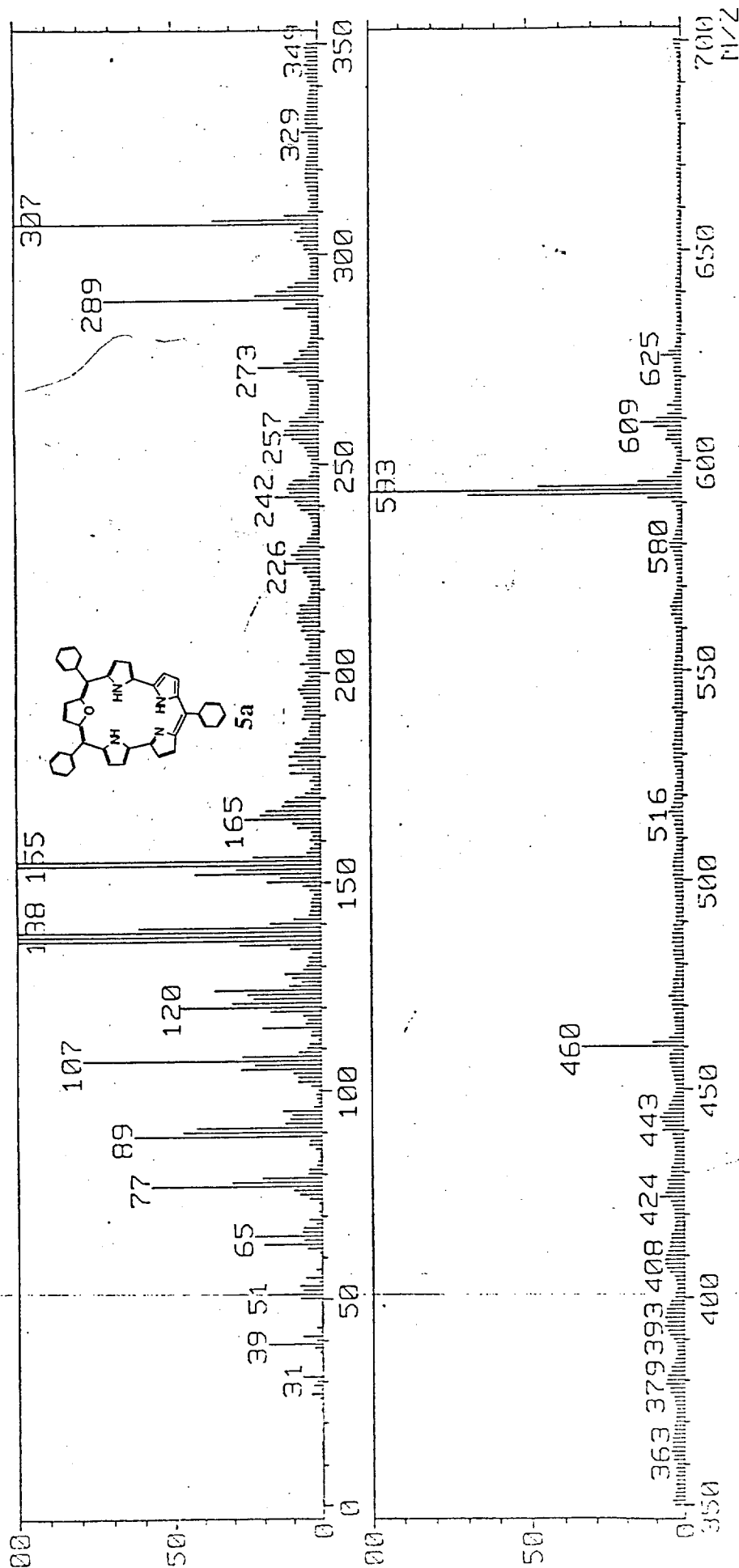
2D-COSY (300 MHz) spectrum of 6a in CDCl₃



2D-Total COSY (300 MHz) spectrum of 6a in CDCl₃.

510

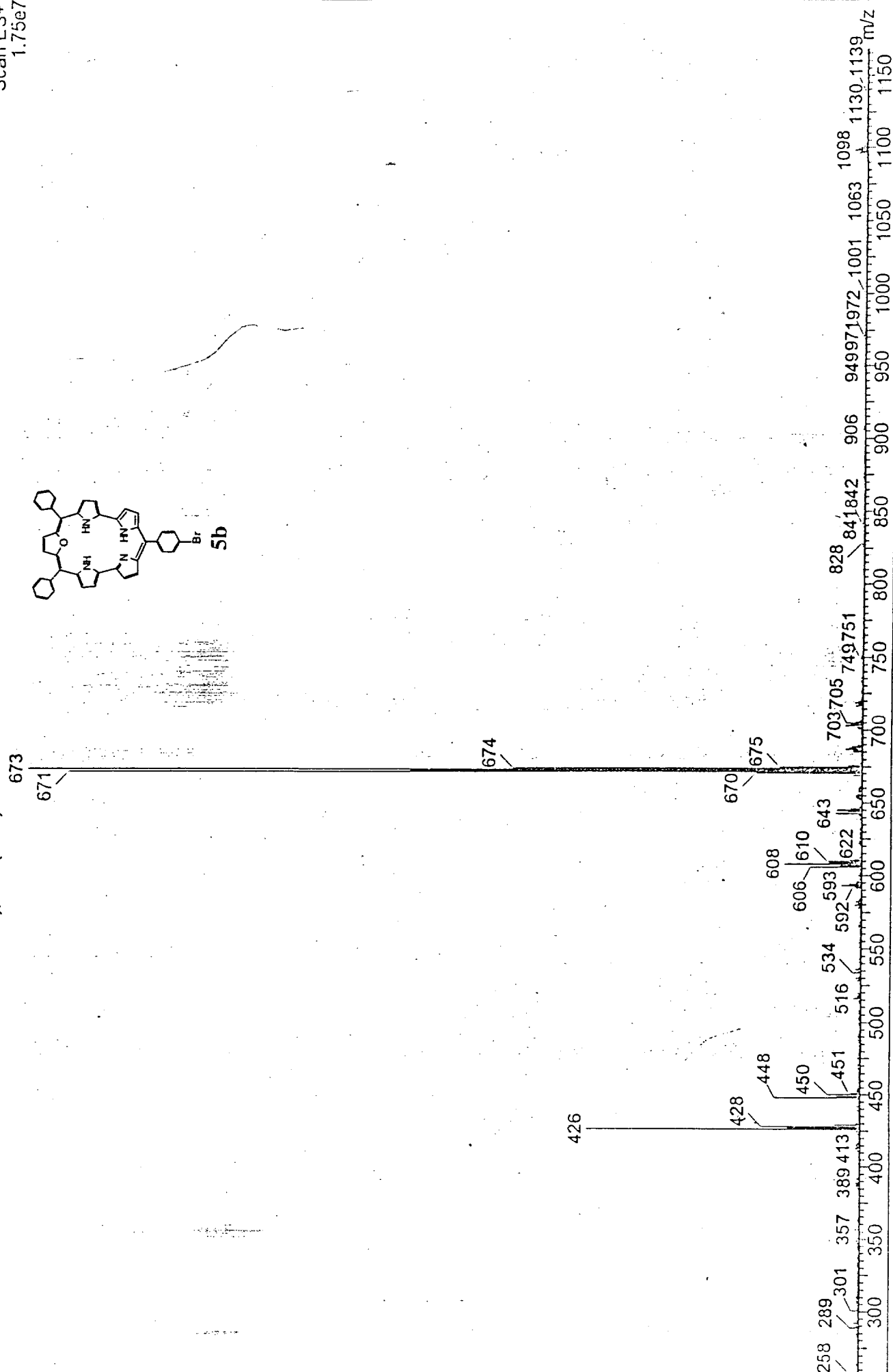
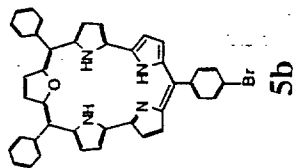
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FAB Mass spectrum of 5a

OSMAR, DR TK CHANDRASEKHAR IITK [382]
 662 1 (0.100) Cn (Cen,2, 80.00, HI); Sm (SG, 2x0.75); Cm (1:10)

Scan ES+
 1.75e7

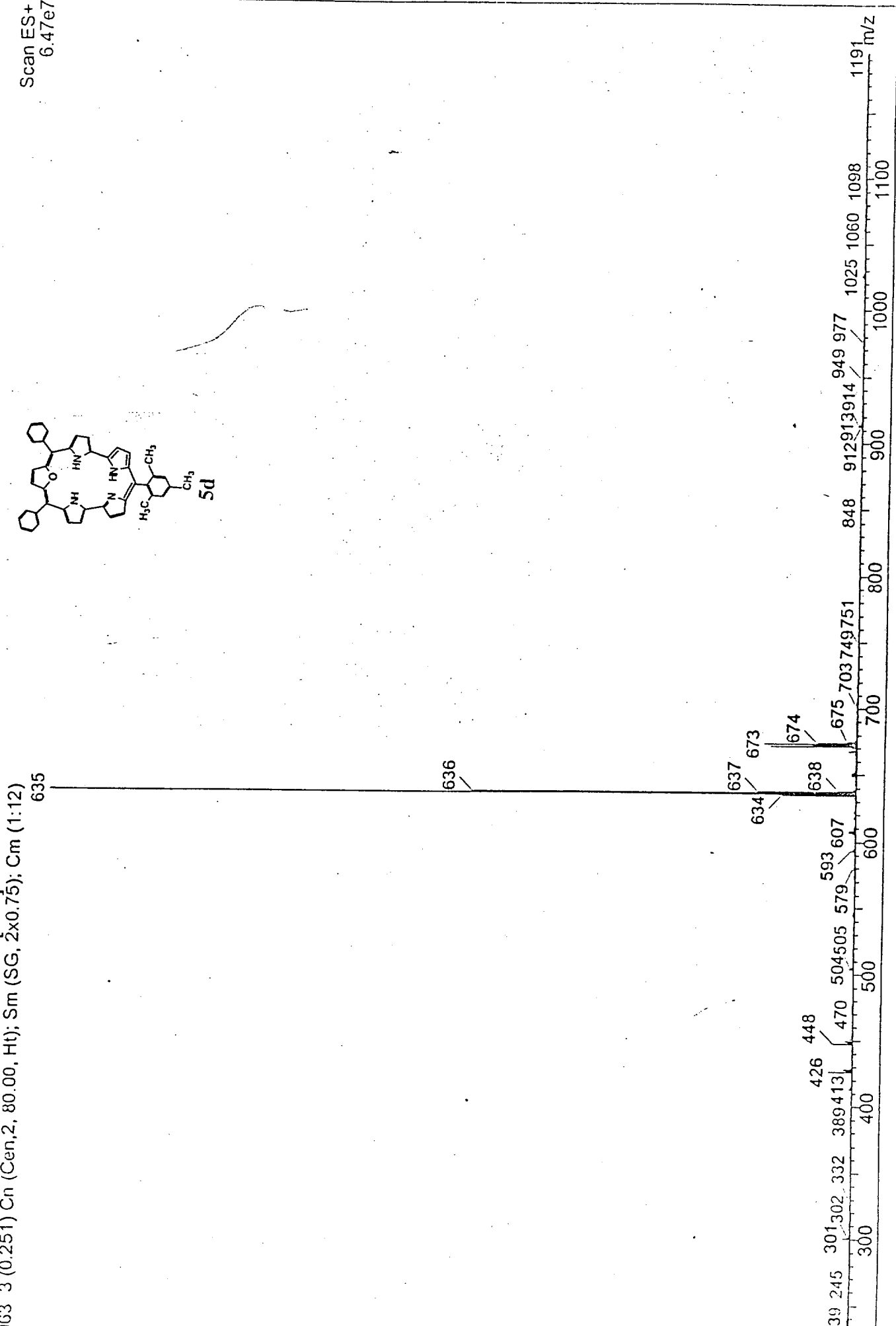
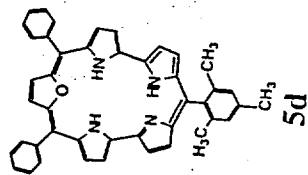


Electron Spray Mass spectrum of 5b

MAR, DR TK CHANDRASEKHAR IITK [382]

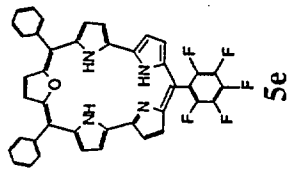
63 3 (0.251) Cn (Cen,2, 80.00, Ht); Sm (SG, 2x0.75); Cm (1:12)

Scan ES+
6.47e7

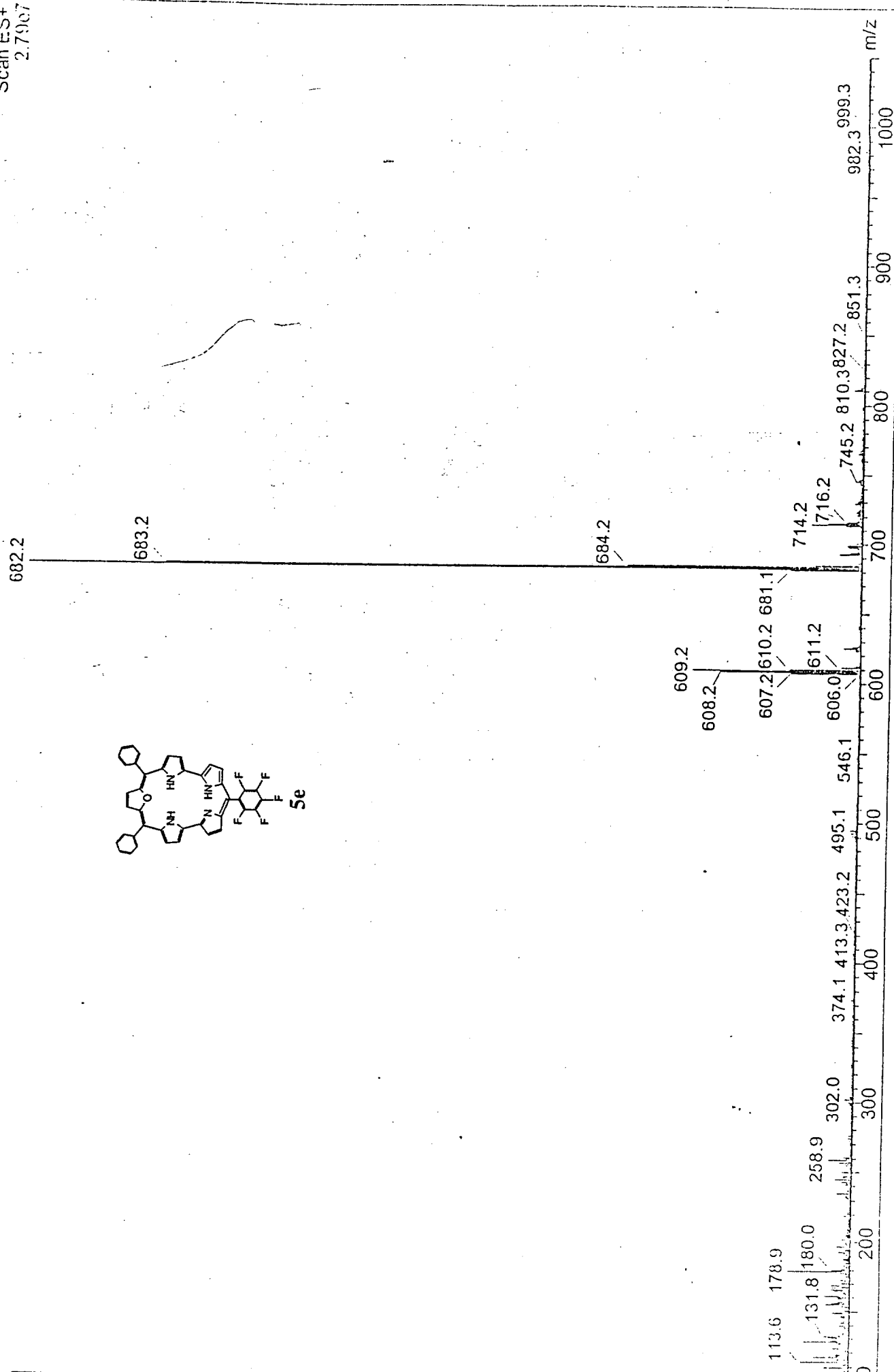


Electron Spray Mass Spectrum of 5d

SMAR, PROF TK CHANDRASHEKAR [638]
 0026E 6 (0.509) Cn (Cen, 2, 80.00, Ht); Sm (SG, 2x0.75); Cm (1:6)

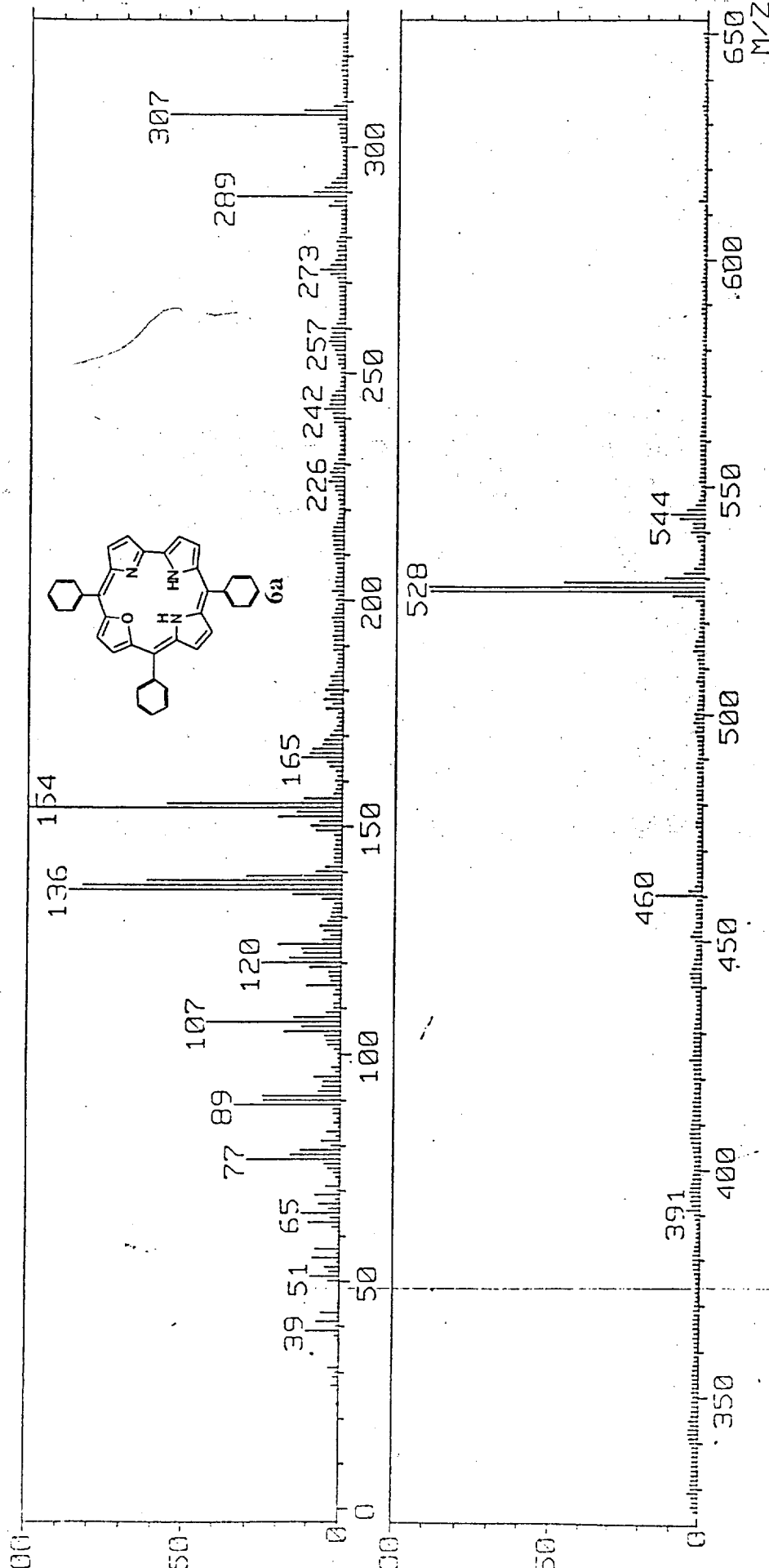


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 2.79e7



Electron Spray Mass spectrum of 5e

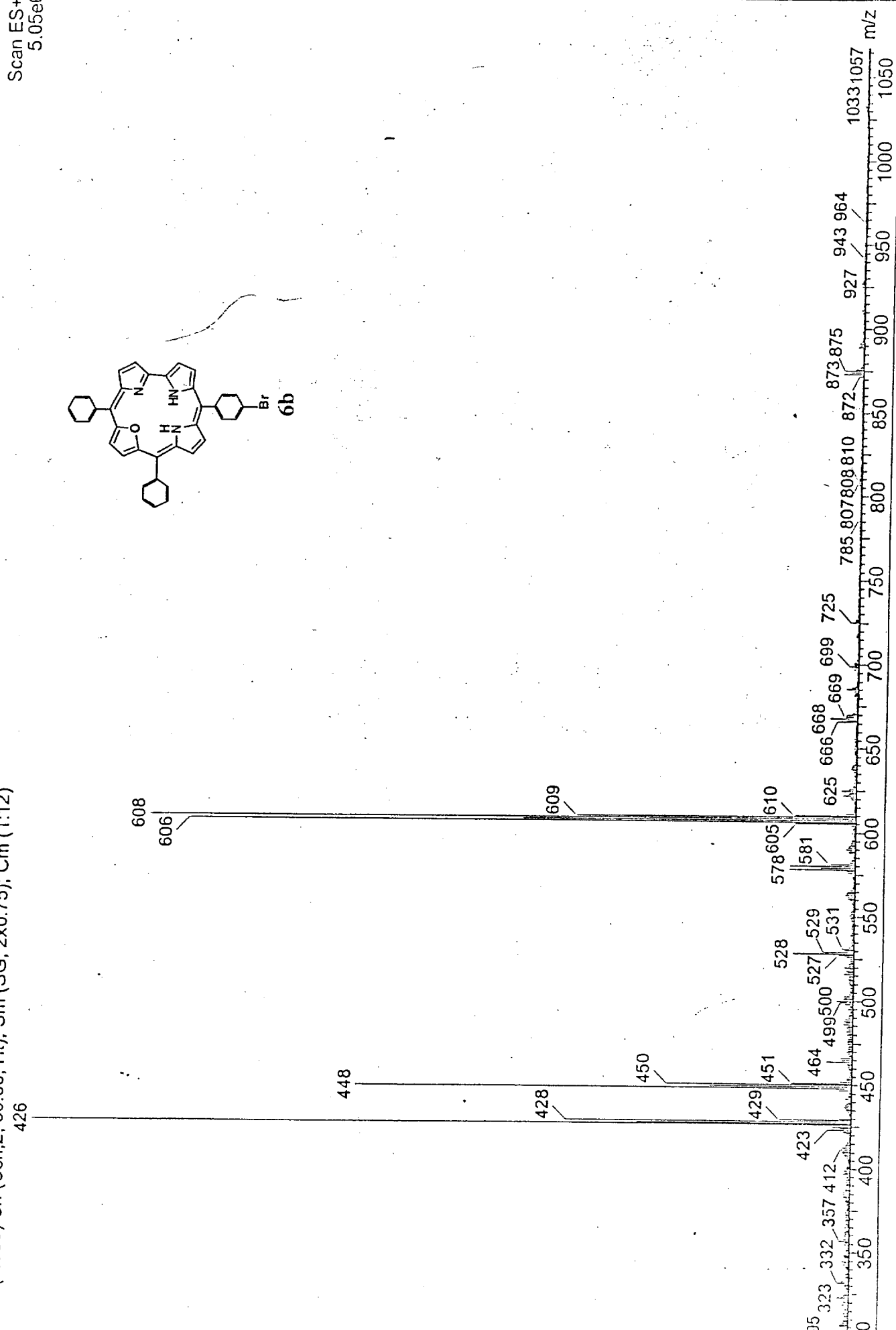
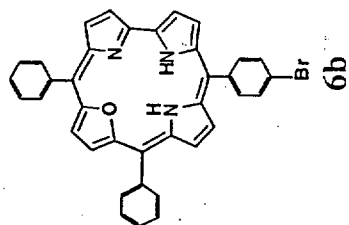
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FAB Mass spectrum of 6a

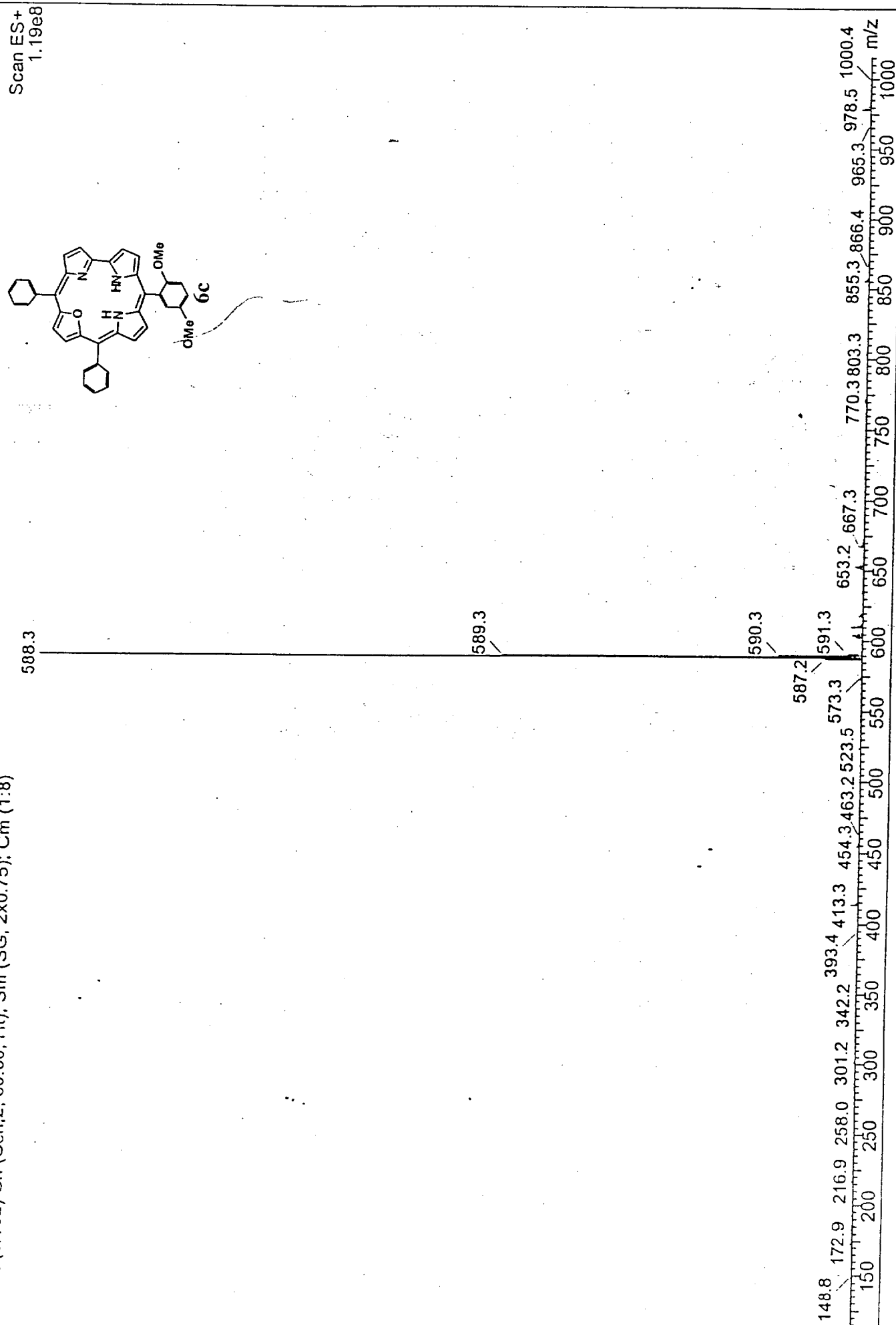
00CORR, DR TK CHANDRASEKHAR IITK [382]
 31 7 (0.550) Cn (Cen,2, 80.00, Ht); Sm (SG, 2x0.75); Cm (1:12)

Scan ES+
 5.05e6



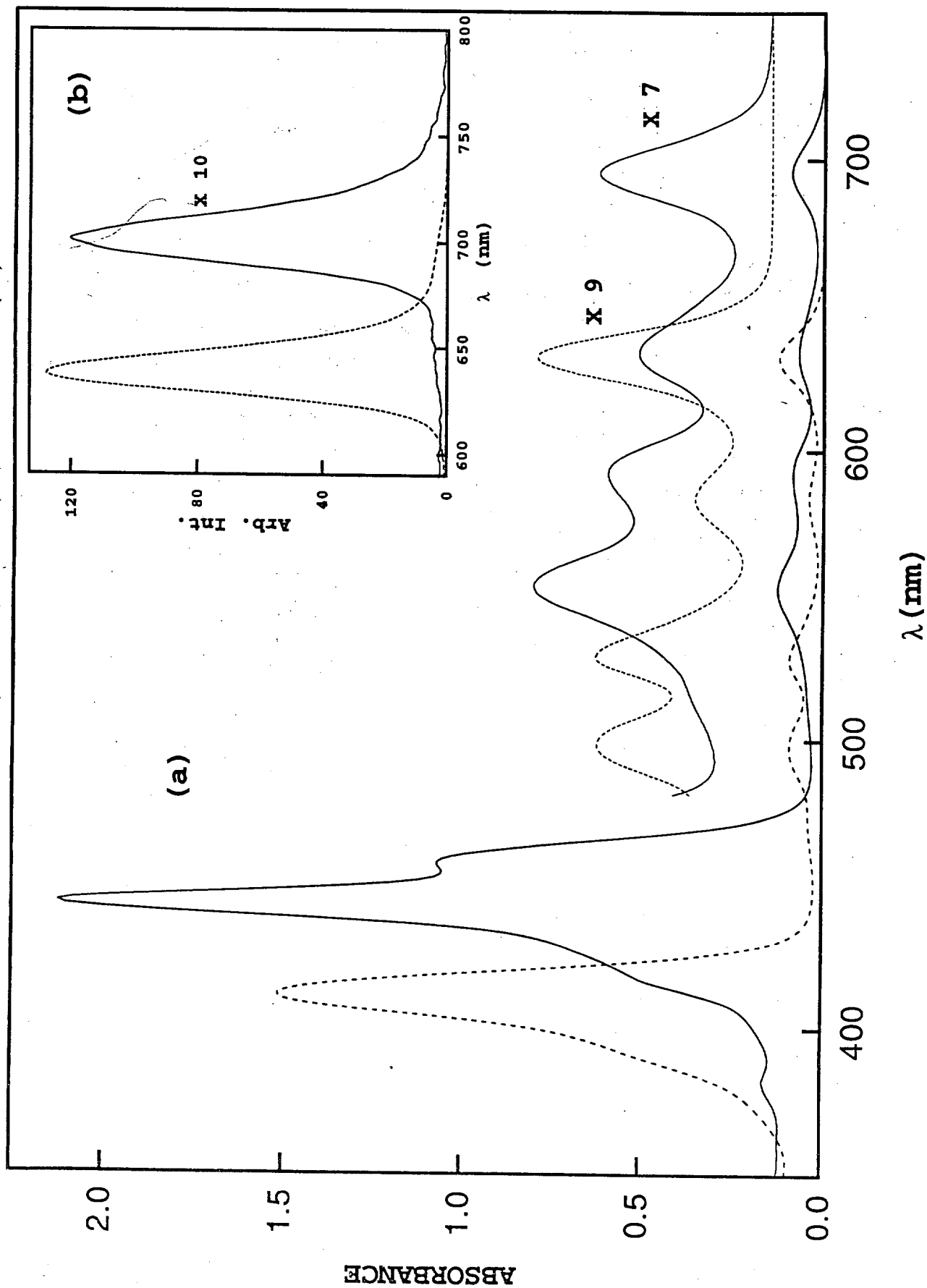
Electron Spray Mass spectrum of 6b

ORR, PROFITK CHANDRASEKAR [603]
 556E 2 (0.192) Cn (Cen, 2, 80.00, H1); Sm (SG, 2x0.75); Cm (1:8)



Electron Spray Mass spectrum of **6c**

The electronic (a) and fluorescence (b) spectra of 5a (solid line) and 6a (dashed line) in CH₂Cl₂. Excitation wavelengths are 443nm (5a) and 411nm (6a).



Selected physical data for the new compounds :

5a: ^1H NMR (300 MHz, CDCl_3 , 25°C, TMS): δ =7.83 (m, 9H), 8.21 (m, 4H), 8.38 (m, 2H), 8.42 (d, 2H, J =4.2Hz), 8.74 (s, 2H), 8.95 (d, 2H, J =4.2 Hz), 9.36 (d, 2H, J =4.2 Hz), 9.46 (d, 2H, J =4.2 Hz); UV/Vis (CH_2Cl_2): λ_{max} ($\text{ex}10^{-4}$); 443 (33.0), 456sh (16.0), 552 (2.0), 591 (1.4), 633 (1.0), 696nm (1.4); UV/Vis ($\text{CH}_2\text{Cl}_2/\text{HCl}$): λ_{max} ($\text{ex}10^{-4}$); 450 (27.9), 482 (13.5), 605 (1.8), 657 (2.4), 720nm (4.6); Emission (CH_2Cl_2): λ =701nm; MS(FAB): m/z (%): 593 (100) [M^+]

5b: ^1H NMR (500 MHz, CDCl_3 , 25°C, TMS): δ =7.82 (m, 6H), 7.99 (d, 2H, J =7.5 Hz), 8.22 (m, 4H), 8.26 (d, 2H, J =7.5Hz), 8.47 (d, 2H, J =4.0Hz), 8.77 (s, 2H), 8.94 (d, 2H, J =4.0 Hz), 9.42 (d, 2H, J =4.0 Hz); 9.52 (d, 2H, J =4.0 Hz); UV/Vis (CH_2Cl_2): λ_{max} ($\text{ex}10^{-4}$); 444 (24.0), 457sh (11.9), 554 (1.46), 594 (1.17), 637 (1.0), 699nm (1.17); UV/Vis ($\text{CH}_2\text{Cl}_2/\text{HCl}$): λ_{max} ($\text{ex}10^{-4}$); 452 (19.6), 483 (8.9), 606 (1.17), 662 (1.84), 724nm (3.5); Emission (CH_2Cl_2): λ =706nm; MS(Electron Spray): m/z (%): 673 (100) [$\text{M}+1$]

5c: ^1H NMR (500 MHz, CDCl_3 , 25°C, TMS): δ =3.67 (s, 3H), 3.97 (s, 3H), 7.71 (s, 1H), 7.80 (m, 8H), 8.22 (m, 4H), 8.44 (d, 2H, J =4.0 Hz), 8.74 (s, 2H), 8.87 (d, 2H, J =4.5 Hz), 9.35 (d, 2H, J =4.0 Hz), 9.48 (d, 2H, J =4.0 Hz); UV/Vis (CH_2Cl_2): λ_{max} ($\text{ex}10^{-4}$); 444 (32.0), 552 (2.6), 592 (2.0), 634 (1.8), 696nm (1.8); UV/Vis ($\text{CH}_2\text{Cl}_2/\text{HCl}$): λ_{max} ($\text{ex}10^{-4}$); 450 (28.0), 479 (16.6), 605 (3.1), 658 (4.4), 712nm (5.9); Emission (CH_2Cl_2): λ =701nm; MS (Electron Spray): m/z (%): 653 (100) [M^+]

5d: ^1H NMR (500 MHz, CDCl_3 , 25°C, TMS): δ =2.04 (s, 3H), 2.64 (s, 6H), 7.31 (s, 2H), 7.80 (m, 6H), 8.20 (m, 4H), 8.38 (d, 2H, J =4.0 Hz), 8.66 (d, 2H, J =4.5 Hz), 8.69 (s, 2H), 9.28 (d, 2H, J =4.0 Hz), 9.42 (d, 2H, J =4.5 Hz); UV/Vis (CH_2Cl_2): λ_{max} ($\text{ex}10^{-4}$); 442 (14.88), 456sh (7.71), 551 (0.88), 590 (0.62), 631 (0.54), 692nm (0.56); UV/Vis ($\text{CH}_2\text{Cl}_2/\text{HCl}$): λ_{max} ($\text{ex}10^{-4}$); 448 (13.09), 477 (6.19), 600 (0.97), 656 (1.38), 707nm (2.24); Emission (CH_2Cl_2): λ =697nm ;MS(FAB) : m/z (%): 635 (100) [M^+]

5e: ^1H NMR (500 MHz, CDCl_3 , 25°C, TMS): δ =-4.1 (s, 1H), 7.86 (m, 6H), 8.28 (m, 4H), 8.71 (d, 2H, J =4.5 Hz), 8.98 (d, 2H, J =4.0 Hz), 9.00 (s, 2H), 9.64 (d, 2H, J =4.0 Hz), 9.75 (d, 2H, J =4.5 Hz); UV/Vis (CH_2Cl_2): λ_{max} ($\text{ex}10^{-4}$); 445 (23.7), 459sh (12.8), 558 (1.30), 600 (1.20), 645 (0.90), 710nm (1.65); UV/Vis ($\text{CH}_2\text{Cl}_2/\text{HCl}$): λ_{max} ($\text{ex}10^{-4}$); 450 (20.6), 481 (9.9), 611 (1.3), 667 (2.2), 714nm (3.6); Emission (CH_2Cl_2): λ =716nm; MS(Electron Spray): m/z (%): 682 (100) [M^+].

5f: ^1H NMR (500 MHz, CDCl_3 , 25°C, TMS): δ =7.85 (m, 9H), 8.32 (m, 4H), 8.35 (m, 2H), 8.63 (d, 2H, J =1.0 Hz), 8.91 (d, 2H, J =4.0 Hz), 9.22 (s, 2H), 9.36 (d, 2H, J =4.0 Hz), 9.38 (d, 2H, J =4.5 Hz); UV/Vis (CH_2Cl_2): λ_{max} ($\text{ex}10^{-4}$); 453 (12.6), 557 (1.11), 590 (0.77), 643 (0.57), 710 (0.46), 823nm (0.19); UV/Vis ($\text{CH}_2\text{Cl}_2/\text{HCl}$): λ_{max} ($\text{ex}10^{-4}$); 478 (10.85), 616 (0.86), 671 (0.86), 878sh (0.95), 927nm (1.34); Emission (CH_2Cl_2): λ =725nm; MS(Electron Spray): m/z (%): 609 (85) [M^+]

6a: ^1H NMR (300 MHz, CDCl_3 , 25°C, TMS): δ =-1.91 (s, 1H), 7.77 (m, 9H), 8.13 (m, 2H), 8.30 (m, 4H), 8.35 (d, 1H, J =4.5 Hz), 8.57 (q, 1H), 8.78 (d, 1H, J =5.1 Hz), 8.80 (d, 1H, J =4.8 Hz), 8.82 (d, 1H,

J=4.2 Hz), 9.03 (d, 1H, J=4.2 Hz), 9.05 (q, 1H), 9.09 (d, 1H, J=5.1 Hz); UV/Vis (CH₂Cl₂): λ_{\max} (ex10⁻⁴); 411 (27.0), 497 (1.6), 528 (1.6), 583 (0.7), 632nm (2.1); UV/Vis (CH₂Cl₂/HCl): λ_{\max} (ex10⁻⁴); 412 (24.3), 431sh (13.7), 523 (3.2), 583 (0.3), 635nm (5.4); Emission (CH₂Cl₂): λ =640nm; Emission (CH₂Cl₂/HCl): λ =652nm; MS(FAB) : m/z (%): 528 (100) [M⁺]

6b: ¹HNMR (500 MHz, CDCl₃, 25°C, TMS): δ =-2.01 (s, 1H), 7.79 (m, 6H), 7.93 (m, 2H), 8.18 (m, 4H), 8.29 (m, 2H), 8.37 (d, 1H, J=4.5 Hz), 8.59 (q, 1H), 8.76 (d, 1H, J=4.5 Hz), 8.79 (d, 1H, J=3.5 Hz), 8.79 (d, 1H, J=5.0 Hz), 9.05 (d, 1H, J=4.0 Hz), 9.08 (d, 1H, J=5.5 Hz), 9.11 (d, 1H, J=5.5 Hz); UV/Vis (CH₂Cl₂) : λ_{\max} (ex10⁻⁴); 412 (38.0), 498 (2.5), 529 (2.2), 584 (1.1), 633nm (3.6); UV/Vis (CH₂Cl₂/HCl): λ_{\max} (ex10⁻⁴); 414 (36.9), 526 (2.7), 563 (2.5), 586 (2.7), 637nm (7.5); Emission (CH₂Cl₂): λ =641nm; Emission (CH₂Cl₂/HCl): λ =651nm; MS(Electron Spray): m/z (%): 608 (100) [M+1]

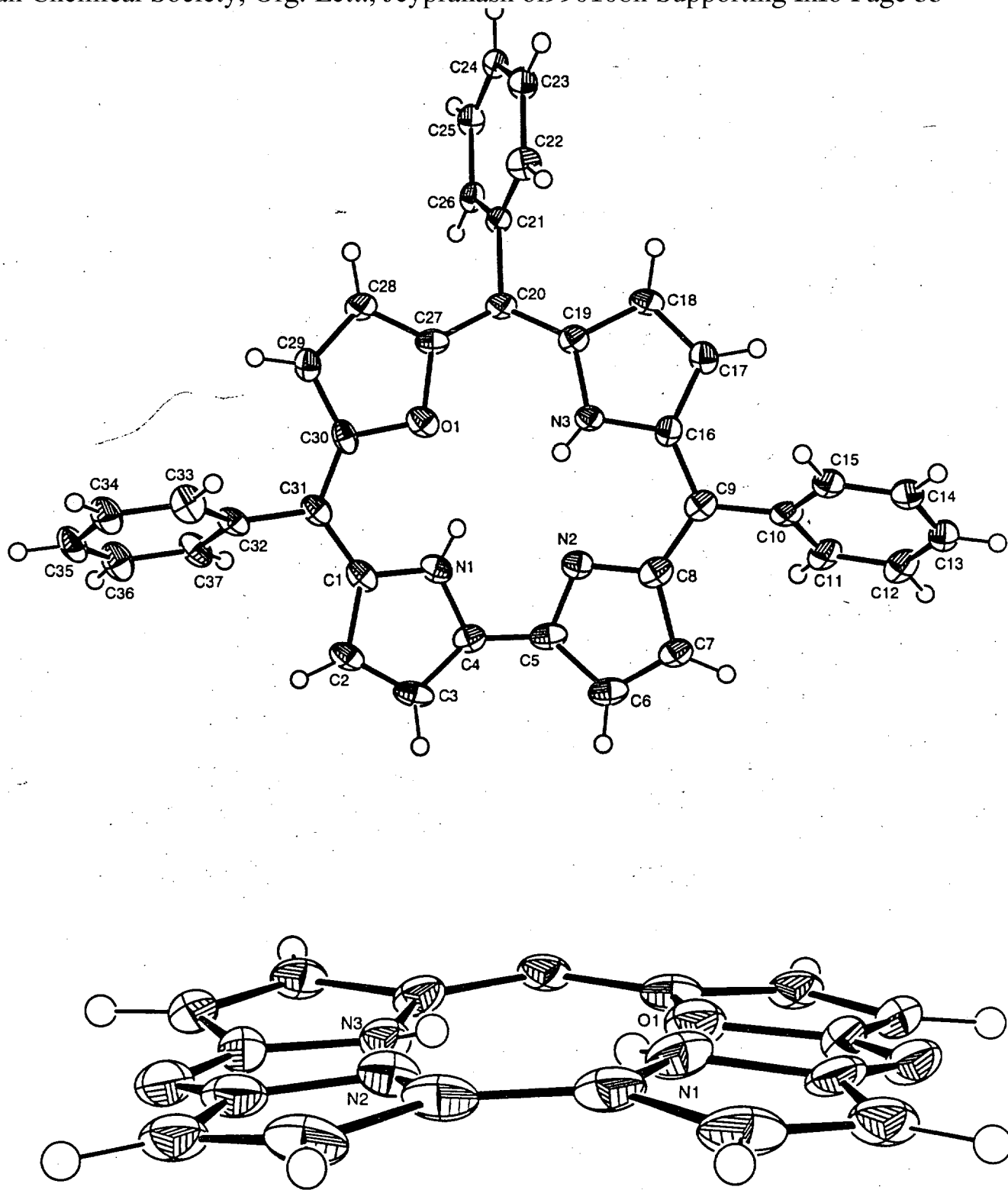
6c: ¹HNMR (500 MHz, CDCl₃, 25°C, TMS): δ =-1.99 (s, 1H), 3.62 (s, 3H), 3.93 (s, 3H), 7.58 (s, 1H), 7.74 (m, 6H), 7.81 (m, 2H), 8.12 (m, 2H), 8.29 (m, 2H), 8.30 (d, 1H, J=4.5 Hz), 8.56 (q, 1H), 8.65 (d, 1H, J=4.5 Hz), 8.73 (d, 1H, J=4.5 Hz), 8.77 (d, 1H, J=5.0 Hz), 8.98 (d, 1H, J=4.0 Hz), 9.05 (q, 1H), 9.08 (d, 1H, J=5.0 Hz); UV/Vis (CH₂Cl₂): λ_{\max} (ex10⁻⁴); 410 (22.65), 498 (1.42), 529 (1.67), 582 (0.64), 630nm (1.94); UV/Vis (CH₂Cl₂/HCl): λ_{\max} (ex10⁻⁴); 411 (15.82), 530 (1.04), 573 (1.19), 629nm (2.61); Emission (CH₂Cl₂): λ =638nm; Emission (CH₂Cl₂/HCl): λ =646nm; MS(Electron Spray): m/z (%): 588 (100) [M⁺]

General synthetic procedure :

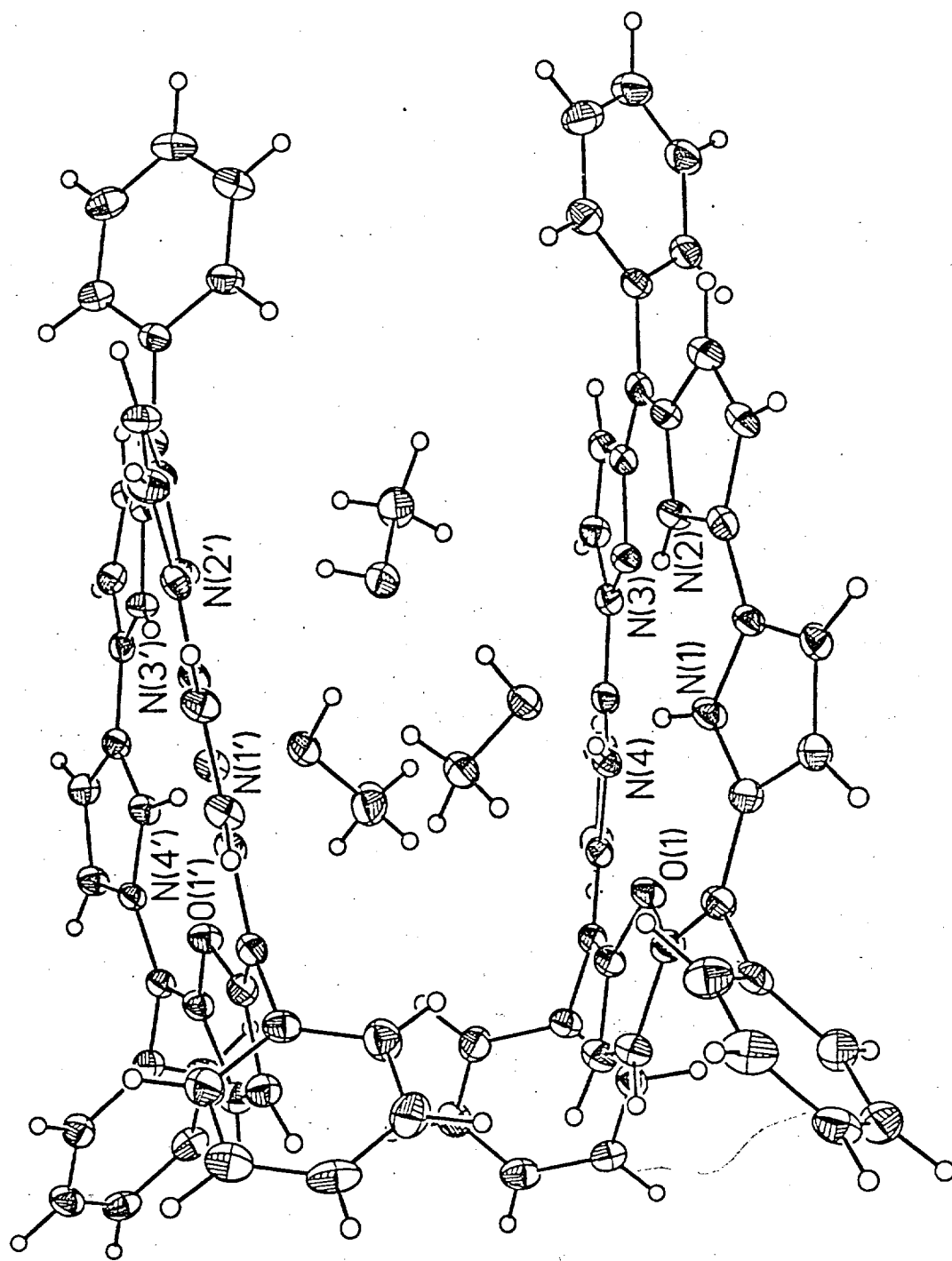
Tripyrrane **3a** (1.263 mmol) and dipyrromethane **4a** (1.263 mmol) was dissolved in 500 ml dry dichloromethane and stirred under nitrogen atmosphere for 5 min. TFA (0.1263 mmol) was added and the stirring continued for further 90 min. Then chloranil (3.789 mmol) was added and the reaction mixture heated at reflux for an additional 90 min. After removal of the solvent the residue was purified by chromatography on basic alumina (grade 3). The pink fraction which eluted with petroleum ether/dichloromethane (5/1) was identified as **6a** (19mg, 3%); smaragdyrin **5a** (380mg, 51%) eluted as a green band when the eluent was dichloromethane. The same procedure was followed for other reactions with appropriate precursors.

Yield of Smaragdyrins and Corroles

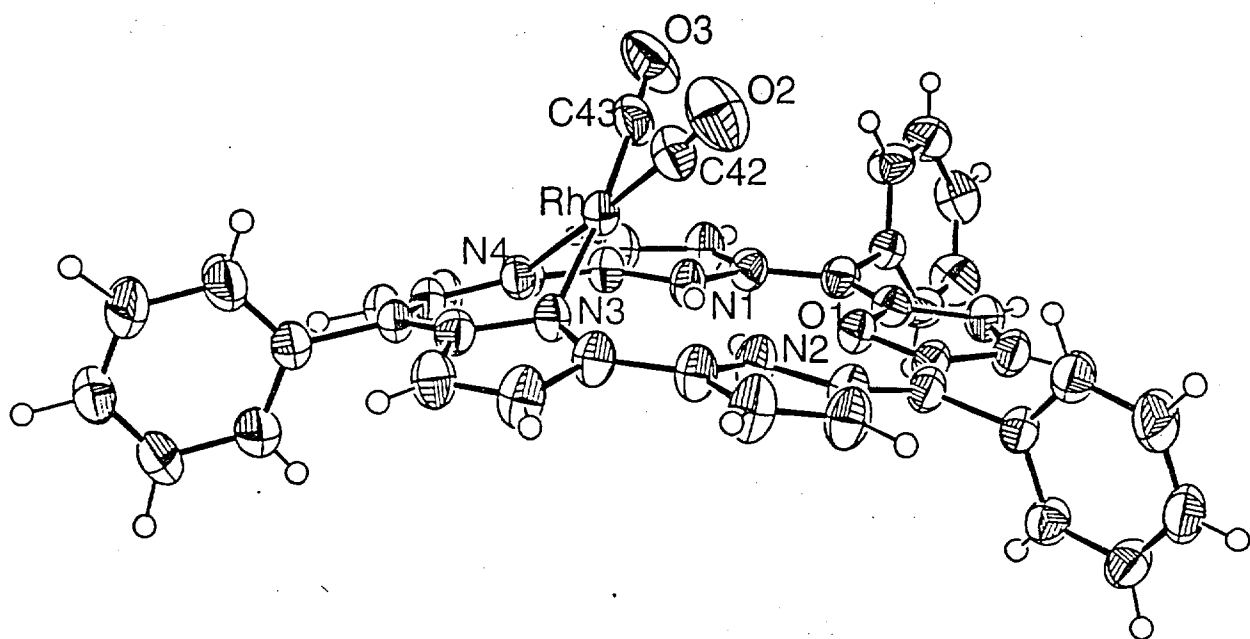
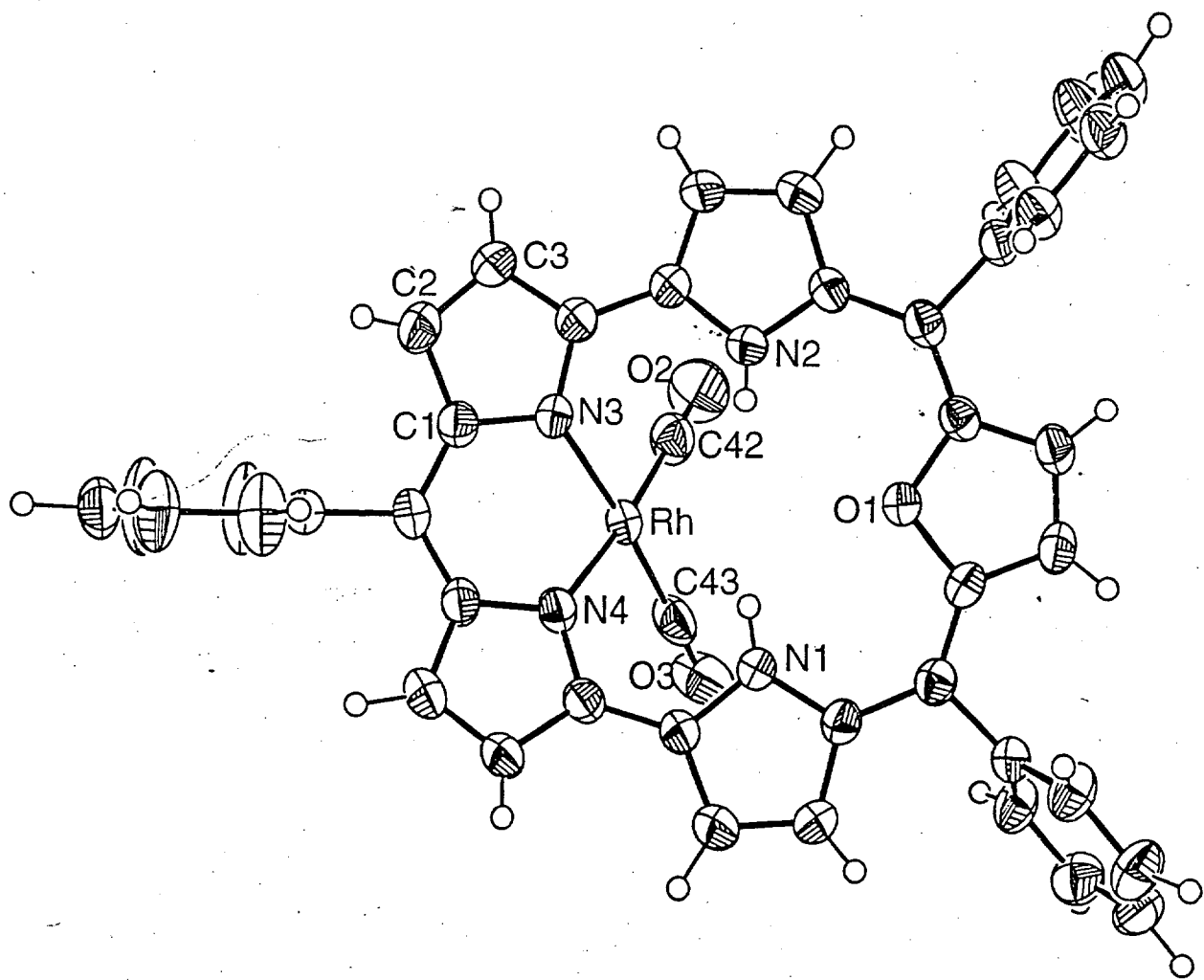
Substituents on the phenyl rings	Equivalents of TFA added	Smaragdyrins		Corroles	
		Compound No.	Yield (%)	Compound No.	Yield (%)
No substituents	0.1	5a	51	6a	3
No substituents	0.5	5a	48	6a	4
No substituents	1	5a	33	6a	6
p-Br	0.5	5b	34	6b	3
2,5- dimethoxy	0.5	5c	13	6c	6
2,4,6-trimethyl (mesityl)	0.3	5d	6	6d	< 1
pentafluoro-	0.5	5e	28	6e	< 1
No substituents	0.1	5f	5	6f	< 1



Crystal structure of Monooxa Corrole. Top: plane view; Bottom: side view (meso phenyl rings are omitted for clarity)



The ORTEP plot showing two independent molecules of 5a trapping three solvent methanol molecules with weak interactions in the unit cell.



ORTEP Plots of Rh(CO)₂ Oxa Smaragdyrin

Selected bond lengths and bond angles :

Rh-N3 2.034, Rh-N4 2.042, Rh-C42 1.871, Rh-C43 1.856, C42-O2 1.126, C43-O3 1.127
N3-Rh-N4 82.45, N4-Rh-C43 94.27, N3-Rh-C42 93.73, C42-Rh-C43 89.32

Table 1. Crystal data and structure refinement for OSMAR.

Identification code	OSMAR
Empirical formula	$C_{85}H_{68}N_8O_5$
Crystal color, habit	deep purple block
Crystal size	0.4 x 0.2 x 0.2 mm
Crystal system	Triclinic
Space group	$P\bar{1}$
Unit cell dimensions	$a = 11.7055(2) \text{ \AA}$ $\alpha = 80.1370(10)^\circ$ $b = 14.83020(10) \text{ \AA}$ $\beta = 79.2780(10)^\circ$ $c = 19.6421(4) \text{ \AA}$ $\gamma = 80.5760(10)^\circ$
Volume	$3269.95(9) \text{ \AA}^3$
Peaks to determine cell	6492 with $I > 10\sigma(I)$
Z	2
Formula weight	1281.47
Density (calculated)	1.302 g/cm^3
Absorption coefficient	0.082 mm^{-1}
F(000)	1348

Data Collection

Diffractionmeter	CCD area detector
θ range for data collection	1.79 to 28.31 $^{\circ}$
Index ranges	-14 $\leq h \leq$ 15, -9 $\leq k \leq$ 19, -26 $\leq l \leq$ 25
Scan Type	phi and omega scans
Scan Time	30sec / frame
Scan Range	0.3 $^{\circ}$ in phi and omega scans
Temperature	98(2) K
Wavelength	0.71073 Å
Detector-to-sample distance	4.956 cm
Reflections collected	21116
Independent reflections	14489 ($R_{int} = 0.0308$)
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.98 and 0.84

The data collection nominally covered over a hemisphere of reciprocal space, by a combination of three sets of exposures; each set had a different ϕ angle for the crystal and each exposure covered 0.3 $^{\circ}$ in ω . The crystal-to-detector distance was 4.956 cm. Coverage of the unique set is over 97% complete to at least 26 $^{\circ}$ in θ . Crystal decay was monitored by repeating the initial frames at the end of data collection and analyzing the duplicate reflections. No decay was observed.

Solution and Refinement

Solution direct methods
 Refinement method Full-matrix least-squares on F^2
 Hydrogen atoms

Positions of all hydrogen atoms were found in difference Fourier maps and the atoms were refined free in their positional and displacement parameters. No constraints were applied.

Weighting scheme

$$w = 1/[\sigma^2(F_o^2) + (0.0574P)^2 + 0.0P]$$

$$\text{where } P = [F_o^2 + 2F_c^2]/3$$

Data / restraints / parameters 14489 / 0 / 1156
 Goodness-of-fit on F^2 0.946
 Final R indices [$I > 2\sigma(I)$] R1 = 0.0541, WR2 = 0.1034
 R indices (all data) R1 = 0.1134, WR2 = 0.1232
 Observed data [$I > 2\sigma(I)$] 8571
 Extinction coefficient 0.0024(3)
 Largest diff. peak and hole 0.337 and -0.298 $e\text{\AA}^{-3}$
 Largest and mean Δ / esd 0.001 and 0.000

$$WR2 = [\sum[w(F_o^2 - F_c^2)^2] / \sum[w(F_o^2)^2]]^{1/2}$$

$$R1 = \sum||F_o| - |F_c|| / \sum|F_o|$$

Data Collection:

SMART Software Reference Manual (1994). Siemens Analytical
X-ray Instruments, 6300 Enterprise Dr., Madison, WI 53719-1173, USA.

Data Reduction:

SAINT Version 4 Software Reference Manual (1995). Siemens Analytical
X-ray Instruments, 6300 Enterprise Dr., Madison, WI 53719-1173, USA.

Structure Solution, Refinement and Graphics:

G. M. Sheldrick (1994). SHELXTL Version 5 Reference Manual. Siemens Ana-
lytical X-ray Instruments, 6300 Enterprise Dr., Madison, WI 53719-1173, USA.

Neutral atom scattering factors were taken from:

International Tables for Crystallography, Vol C, Tables 6.1.1.4,
4.2.6.8, and 4.2.4.2, Kluwer: Boston.

Acknowledgement

Please acknowledge funds from NSF (grant CHE-9527898), Syracuse
University, and the W.M. Keck Foundation for the purchase of
the X-ray instrument and computers.

Table 2. Atomic coordinates [$\times 10^4$] and equivalent isotropic displacement parameters [$\text{\AA}^2 \times 10^3$] for OSMAR. $U(\text{eq})$ is defined as one third of the trace of the orthogonalized U_{ij} tensor.

	x	y	z	U(eq)
O(1)	3435(1)	1868(1)	1772(1)	21(1)
N(1)	4363(2)	1152(1)	3114(1)	20(1)
N(2)	3432(2)	2194(1)	4302(1)	20(1)
N(3)	2213(1)	3697(1)	3797(1)	21(1)
N(4)	2096(2)	3606(1)	2355(1)	20(1)
C(1)	5044(2)	588(1)	2661(1)	20(1)
C(2)	5981(2)	113(2)	2998(1)	23(1)
C(3)	5834(2)	385(2)	3652(1)	22(1)
C(4)	4815(2)	1017(1)	3730(1)	19(1)
C(5)	4292(2)	1463(1)	4320(1)	19(1)
C(6)	4557(2)	1265(2)	5008(1)	22(1)
C(7)	3852(2)	1895(2)	5390(1)	23(1)
C(8)	3153(2)	2512(1)	4944(1)	20(1)
C(9)	2449(2)	3367(1)	5026(1)	20(1)
C(10)	2234(2)	3665(2)	5738(1)	21(1)
C(11)	1807(2)	3082(2)	6330(1)	27(1)
C(12)	1621(2)	3358(2)	6990(1)	33(1)
C(13)	1857(2)	4216(2)	7062(1)	34(1)
C(14)	2280(2)	4803(2)	6478(1)	29(1)
C(15)	2473(2)	4527(2)	5817(1)	24(1)
C(16)	2019(2)	3953(1)	4462(1)	20(1)
C(17)	1340(2)	4856(2)	4450(1)	22(1)
C(18)	1142(2)	5146(2)	3783(1)	22(1)
C(19)	1695(2)	4412(2)	3390(1)	21(1)
C(20)	1699(2)	4389(2)	2664(1)	21(1)
C(21)	1345(2)	5092(2)	2149(1)	24(1)
C(22)	1502(2)	4721(2)	1530(1)	24(1)
C(23)	1961(2)	3788(2)	1656(1)	19(1)
C(24)	2198(2)	3138(2)	1175(1)	20(1)
C(25)	1737(2)	3472(1)	495(1)	20(1)
C(26)	593(2)	3397(2)	432(1)	30(1)
C(27)	185(2)	3717(2)	-199(1)	32(1)
C(28)	913(2)	4110(2)	-773(1)	28(1)
C(29)	2053(2)	4184(2)	-718(1)	25(1)
C(30)	2460(2)	3869(2)	-84(1)	22(1)
C(31)	2812(2)	2267(2)	1222(1)	22(1)
C(32)	3002(2)	1631(2)	740(1)	24(1)
C(33)	3745(2)	884(2)	961(1)	23(1)
C(34)	4045(2)	1023(1)	1599(1)	20(1)
C(35)	4826(2)	478(1)	1994(1)	21(1)
C(36)	5572(2)	-314(2)	1675(1)	22(1)
C(37)	6474(2)	-144(2)	1119(1)	28(1)
C(38)	7220(2)	-864(2)	841(1)	30(1)
C(39)	7074(2)	-1768(2)	1110(1)	29(1)
C(40)	6178(2)	-1947(2)	1660(1)	34(1)
C(41)	5433(2)	-1229(2)	1946(1)	30(1)